

Algorithms to estimate Antarctic sea ice algal biomass from under-ice irradiance spectra at regional scales

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ABSTRACT: The presence of algal pigments in sea ice alters under-ice irradiance spectra, and the relationship between these variables can be used as a non-invasive means for estimating ice-associated algal biomass on ecologically relevant spatial and temporal scales. While the influence of snow cover and ice algal biomass on spectra transmitted through the snow-ice matrix has been examined for the Arctic, it has not been tested for Antarctic sea ice at regional scales. We used paired measurements of sea ice core chl *a* concentrations and hyperspectral-transmitted under-ice irradiances from 59 sites sampled off East Antarctica and in the Weddell Sea to develop algorithms for estimating algal biomass in Antarctic pack ice. We compared 4 approaches that have been used in various bio-optical studies for marine systems: normalised difference indices, ratios of spectral irradiance, scaled band area and empirical orthogonal functions. The percentage of variance explained by these models ranged from 38 to 79%, with the best-performing approach being normalised difference indices. Given the low concentrations of integrated chl *a* observed in our study compared with previous studies, our statistical models performed surprisingly well in explaining variability in these concentrations. Our findings provide a basis for future work to develop methods for non-invasive time series measurements and medium- to large-scale spatial mapping of Antarctic ice algal biomass using instrumented underwater vehicles.

KEY WORDS: Sea ice algae · Chl *a* · Bio-optics · Normalised difference index · Weddell Sea · East Antarctica

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INTRODUCTION

Ice algal communities in Antarctic sea ice form an important component of primary production in the Southern Ocean (Lizotte 2001, Arrigo et al. 2010); they support higher trophic levels and act as a food source for overwintering Antarctic krill larvae (Flores et al. 2012, Arrigo 2014). Primary productivity in sea

ice is also critical in large-scale biogeochemical cycles that determine rates of carbon export and atmosphere-ocean exchange (Vancoppenolle et al. 2013). However, the high spatial variability of ice algal distribution prevents accurate *in situ* estimation of biomass at scales that are ecologically relevant—e.g. for regional-scale primary and secondary productivity—using classical sampling designs such as

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ice coring (Meiners et al. 2012). Recent circumpolar estimates of ice algal biomass for Antarctica are limited to records from historical ice core data that are unevenly distributed in space and time (Meiners et al. 2012) and model-based estimates that may underestimate internal communities (Saenz & Arrigo 2014). Improved *in situ* data on temporal and spatial patterns of ice algal biomass distribution are needed for quantitative evaluation of sea ice primary production models and an improved understanding of the role of ice algae in Antarctic marine ecosystem function.

Antarctic pack ice provides a habitat for ice-associated algae, which form distinct surface, interior and bottom communities (Arrigo et al. 2010, Meiners et al. 2012). Surface communities are promoted by snow loading, surface flooding by seawater and brine, and subsequent snow-ice formation (Ackley et al. 2008). Interior communities can form either through the rafting and ridging of ice floes or by scavenging of phytoplankton during ice formation (i.e. the uptake of algal cells from the water column as ice crystals form; Arrigo et al. 2010). Bottom communities thrive in the lowermost porous parts of sea ice floes, where brine salinities and high nutrient availability favour algal growth (e.g. Vancoppenolle et al. 2010). The presence of photosynthetic pigments in these ice algal communities alters under-ice irradiance spectra through absorption at specific wavelengths of photosynthetically active radiation (Arrigo et al. 1993, Perovich et al. 1993, Mundy et al. 2007, Fritsen et al. 2011, Hawes et al. 2012). With increasing ice algal pigment concentration, transmitted under-ice irradiance spectra change from showing a broad peak centred at approximately 460 nm towards a narrow peak at wavelengths of approximately 570 nm. Such a relationship—often measured as the ratio between intensities at key wavelengths—can potentially be used as an alternative non-invasive means for estimating ice-associated algal biomass (e.g. from transmitted spectra measured by instrumented underwater vehicles such as remotely operated vehicles and autonomous underwater vehicles). Several studies examining this relationship for ice algal communities have occurred in the Arctic (e.g. Legendre & Gosselin 1991, Perovich et al. 1993, Mundy et al. 2007, McDonald et al. 2015) and a single study in the Antarctic (Fritsen et al. 2011); however, no study has yet tested the relationship across regional scales.

Transmitted and reflected spectra have been used in the Arctic and other marine environments to predict the concentration of photosynthetic pigments. Legendre & Gosselin (1991) found that the under-ice

transmitted irradiance ratio 671:540 nm explained approximately 50 % of the observed integrated chl *a* concentration in Arctic fast ice. Perovich et al. (1993) demonstrated that the ratio of transmitted light at 600 and 450 nm can be used as an indicator of algal biomass in first-year Arctic sea ice. Mundy et al. (2007) employed normalised difference indices (NDIs) of under-ice irradiance spectra to investigate the influence of both ice algal biomass and snow on transmitted spectra under Arctic fast ice. The authors showed that snow has little effect on transmitted under-ice irradiances in the wavelength range of 400 to 570 nm and that ice algal biomass in Arctic fast ice explained 89 % of the NDI wavelength combination 485:472 nm. Fritsen et al. (2011) used the NDI wavelength combination 555:442 nm to estimate temporal changes in integrated algal biomass at 3 sites in the Bellingshausen Sea (pack ice) during a time series experiment. This NDI wavelength pair explained 81 % of the variation in integrated ice algal biomass.

Other bio-optical studies for open water systems have used simple ratios of spectral irradiance at particular wavelengths to estimate algal biomass. For example, Honda et al. (2009) found that the ratio of spectral irradiance at wavelengths of 555 and 443 nm at 40 m depth (the depth at which a moored radiometer was deployed) explained 80 % of the variation in integrated chl *a* in the surface water column in the Western Pacific Subarctic Gyre. Carrere et al. (2004) compared a series of methods for estimating the concentration of microphytobenthos chl *a* from sediment optical properties. These authors report that scaled band area, calculated by integrating the scaled absorption band depth over the wavelengths of particular absorption features, gave the best predictions of chl *a* ($R^2 = 0.85$) among the methods evaluated in their study. Finally, Craig et al. (2012) and Taylor et al. (2013) use empirical orthogonal function (EOF) analysis to reduce the dimensionality of hyperspectral data and relate independent modes of variability to phytoplankton pigment concentrations. These various approaches have never been directly compared, and it is unclear which might be most effective for predicting algal biomass in Antarctic sea ice.

The aim of our study was to compare existing bio-optical approaches to predict chl *a* concentration in Antarctic pack ice using paired measurements of sea ice core chl *a* concentrations and hyperspectral-transmitted under-ice irradiances from 59 sites sampled off East Antarctica and in the Weddell Sea. Specifically, we compared algorithms based on 4 classes of methods (summarised in Table 1) that have been used for Arctic sea ice and other marine sys-

Table 1. Summary of existing approaches for estimating chl *a* from measurements of transmitted irradiance spectra in marine bio-optical studies. Previous studies ('Reference' in table) used linear models to relate chl *a* (dependent variable) to ratios, indices, scaled areas or modes of oscillation (independent variables) from transmitted irradiance spectra measurements. Formulations of approaches are provided in 'Materials and methods: Analysis of transmitted spectra'. NDI: normalised difference index; EOF: empirical orthogonal function

Approach	Bio-optical relationship with algal pigments	Previous applications	Reference	Input data (this study)
NDIs	As for spectral ratios (see next approach), but the normalised value range (-1 to 1) facilitates inter-comparison. Previous studies suggest a more linear dependence on ice algal biomass.	Arctic fast ice; Bellingshausen Sea, pack ice, time series	Mundy et al. (2007) Fritsen et al. (2011)	Transmitted spectra ^a
Ratios of spectral irradiance	Based on spectral transmittance or transmitted irradiance. Capture absorption peaks for targeted algal pigments.	Western Pacific Subarctic Gyre; Bellingshausen Sea, pack ice	Honda et al. (2009) Fritsen et al. (2011)	Integral-normalised transmitted spectra
Scaled band area	Developed for reflectance curves to capture the strength of absorption features. Integration over absorption features is intended to average out fluctuations caused by noise and produce a more realistic estimate of absorption strength.	Microphytobenthos (sediment optical properties)	Carrere et al. (2004)	Integral-normalised transmitted spectra
EOF analysis	Reduces dimensionality of the data (i.e. number of wavelengths) and results in a set of orthogonal modes of oscillation that can be correlated with algal biomass.	Phytoplankton	Craig et al. (2012) Taylor et al. (2013)	Integral-normalised transmitted spectra

^aNormalisation is achieved through the NDI algorithm

tems: NDIs, simple ratios of spectral irradiance, integrated features of transmitted irradiance spectra (scaled band areas) and EOF analysis. We directly compared the predictive capability of these approaches and examined their applicability between the 2 separate regions of Antarctica.

MATERIALS AND METHODS

Study sites

Paired under-ice irradiance measurements and ice core samples were collected during 3 expeditions (Fig. 1): the Winter Weddell Outflow Study (WWOS) voyage onboard the RV 'Polarstern' to the Weddell Sea in austral spring (September to October) 2006 (Lemke 2009) and 2 Sea Ice Physics and Ecosystems Experiment voyages (SIPEX1 and SIPEX2) onboard the RV 'Aurora Australis' to the 115 to 130°E sector off East Antarctica in austral spring (September to October) 2007 and 2012 (Worby et al. 2011). For the purposes of our analyses, i.e. our focus on developing and comparing algorithms between regions, data from the 2 SIPEX voyages in East Antarctic are combined. Exploratory analyses (model fitting) indicated that statistical models were more robust (less prone to overfitting) with the larger combined data set for East Antarctica.

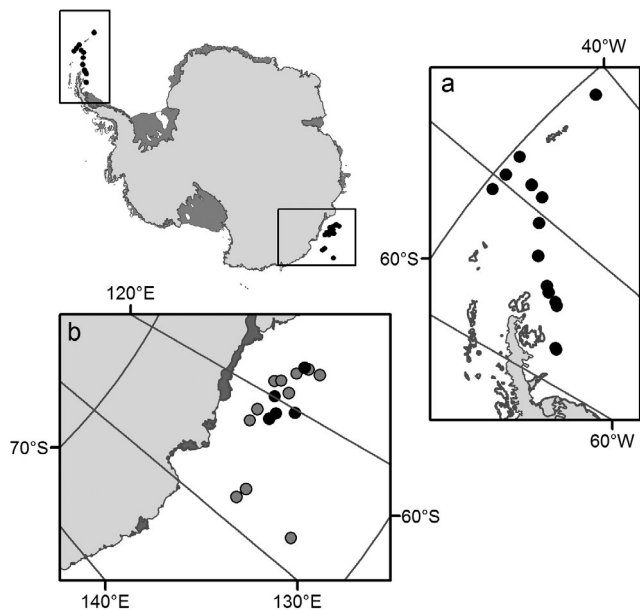


Fig. 1. Survey locations in (a) the Weddell Sea and (b) off East Antarctica. Station locations for the Winter Weddell Outflow Study (WWOS) voyage (2006) are shown in (a), and station locations for Sea Ice Physics and Ecosystems Experiments 1 (SIPEX1; 2007, ●) and 2 (SIPEX2; 2012, ●) are shown in (b)

Antarctic sea ice extent is at its maximum in September (Parkinson & Cavalieri 2012). Sea ice in the areas visited during the 3 voyages considered here was snow covered and was generally beginning to thin and loosen (break up) in the September to October period. Sampled ice floes on these voyages were several hundred meters to several kilometres in diameter and were all composite sea ice floes showing different degrees of deformation (ridging and rafting). Bio-optical sampling sites (1 to 5 sites per ice station, separated from each other by at least 20 m) were chosen such that their immediate surroundings (20 m) showed no snow cover disturbances, were level and were free of surface deformations. All samples were taken during the middle of the day when the solar angle was high.

Spectral measurements

On each sampling site, we deployed a TriOS Ramesses ACC VIS radiometer (cosine receptor, 180° field of view) 0.15 m beneath the subsurface of the ice floe through an access hole (0.11 m diameter) using a retractable L-shaped stainless steel arm (Fig. 2). The hole effect on the light measurements was assumed to be negligible due to the position of the radiometer 0.9 m north (i.e. directed towards the sun) of the access hole. Under-ice high-resolution (400 to 700 nm, 3.3 nm bandwidth) irradiance spectra were recorded with a laptop computer using TriOS MSDA software version 7.5.1. Results reported here represent average spectra calculated from a minimum of 10 replicate radiometer scans. Note that we measured transmitted irradiance spectra, not transmittance.

Physical measurements and ice core processing

At each site, snow depth was measured with a ruler (5 replicate measurements) and an ice core was collected, using a manual Kovacs Mark II ice core system (0.09 m internal diameter), directly above the radiometer location. Medium-scale spatial variability was addressed by sampling from multiple sites at each ice station.

The length of the collected ice core was recorded, and the entire core was divided into sections (one 10 cm bottom section and 3 equal-length sections) and then placed in clean polyethylene containers and transported back to the ship's laboratories. On the ship, ice core sections were melted at 4°C in the dark within 12 to 36 h of collection. After the ice cores

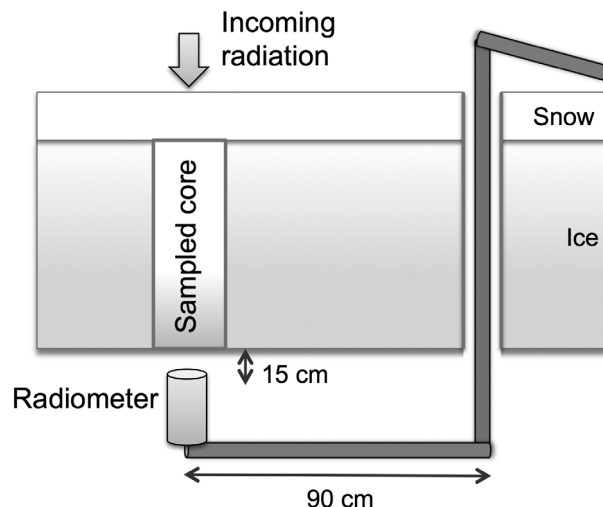


Fig. 2. Sampling design for spectral measurements used on voyages in the Weddell Sea and East Antarctica

had melted, samples were gently mixed, and subsamples were filtered onto Whatman GF/F glass fibre filters, extracted with 90% acetone (WWOS) or methanol (SIPEX voyages) and analysed for chl *a* with a Turner Designs 10AU fluorometer according to standard protocols (Holm-Hansen et al. 1965, Arar & Collins 1997). Ice core section data (chl *a* in $\mu\text{g l}^{-1}$) were integrated over the entire ice thickness and are expressed as integrated chl *a* (mg m^{-2}).

Analysis of transmitted spectra

All transmitted spectra were interpolated to consecutive integer (1 nm) wavelengths in the range of 400 to 700 nm. Ratios, scaled band areas and EOFs were calculated using integral-normalised transmitted irradiance (i.e. normalised E_d over the 400 to 700 nm range) to minimise the amplitude component of spectral variability and to focus on differences in spectral shape. We considered the relationship between snow depth and transmitted irradiance using NDIs but not ratios, scaled band area or EOFs. Mundy et al. (2007) showed that for the Arctic, snow has a minor influence on the spectral distribution of transmitted under-ice irradiances in the wavelength range of 400 to 570 nm. The calculations of ratios and scaled band area described below are therefore unlikely to be influenced by snow depth. Furthermore, because EOFs partition variability in spectra due to different signals, this approach should be capable of partitioning variability due to snow depth as compared with algal biomass (provided that algal biomass is not correlated with snow depth; see 'Algorithms', below).

NDIs

Following Mundy et al. (2007), we calculated NDIs for all possible wavelength combinations of measured transmitted irradiances ($T\lambda$, 400 to 700 nm) according to:

$$\text{NDI} = [(T\lambda_1) - (T\lambda_2)] / [(T\lambda_1) + (T\lambda_2)] \quad (1)$$

The NDIs for each wavelength pair were then correlated with integrated chl *a* and snow depth, and the resultant Pearson correlation coefficients were placed in 2 separate matrices with λ_1 and λ_2 as their axes. NDI wavelength pairs that gave the best correlation with each variable were determined by selecting a threshold between 0.7 and 0.9 that resulted in a single region of correlations higher than the threshold, recording the coordinates of this region and then taking the mid-point.

Ratios

We adopted the wavelength ratio used by Honda et al. (2009) of $E_d(555)/E_d(443)$, noting that this is consistent with the ratio considered by Fritsen et al. (2011; for the Bellingshausen Sea, Antarctica) of $E_d(555)/E_d(442)$.

Scaled band area

Carrere et al. (2004) determined the scaled area of absorption features using reflectance spectra from

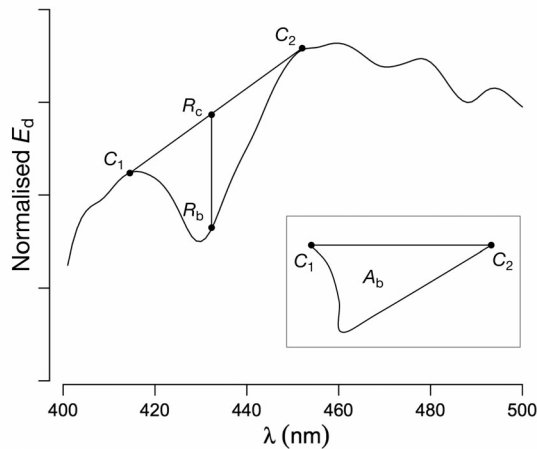


Fig. 3. Calculation of scaled band area (A_b ; Eq. 2) based on normalised spectra for transmitted irradiance. C_1 and C_2 are the wavelengths at which there is maximum transmitted irradiance between 400–430 and 430–450 nm, respectively; R_c is the straight line of irradiance connecting C_1 and C_2 ; and R_b is the normalised spectral irradiance. E_d : transmitted irradiance; λ : wavelength

sediment surfaces in the intertidal zone in the English Channel. Scaled band area is a dimensionless quantity and is calculated by integrating the scaled band depth over the wavelengths of a spectral feature (Fig. 3). We adapted this method to under-ice irradiance spectra and calculated scaled band area, A_b , of the consistent feature of these irradiance spectra between 400 and 450 nm (the interval within which the maximum of chl *a* specific absorption occurs) as:

$$A_b = \int_{C_1}^{C_2} \frac{(R_c - R_b)}{R_c} \quad (2)$$

where C_1 and C_2 are the wavelengths at which there is maximum transmitted irradiance between 400–430 and 430–450 nm, respectively; R_c is the straight line of irradiance connecting C_1 and C_2 ; and R_b is the normalised spectral irradiance.

EOFs

EOF analysis of spectral data has the advantage of reducing the high dimensionality of spectral data and deriving the dominant signals (modes) that best describe variance within the observed spectra. As shown by Craig et al. (2012) and Taylor et al. (2013), EOFs can be represented graphically as independent modes of oscillation in spectral shape.

EOF analysis was conducted using an $M \times N$ matrix of integral-normalised transmitted irradiance at each wavelength (M) for all observations (samples, N). The corresponding covariance matrix was then subjected to an eigen decomposition to obtain eigenvectors (the EOFs) and eigenvalues. The eigenvectors are represented as an $N \times N$ matrix containing scores for each sample by mode. EOF expansion coefficients (i.e. principal components) are represented as an $M \times N$ matrix carrying the loadings for each wavelength by mode. We selected the first 4 EOFs for further analysis.

Algorithms

We used linear models to explore the relationship between integrated chl *a* from ice cores and predictive variables from each method described above (noting that linear models have been used in previous studies estimating chl *a* from bio-optical data). Only cores with ice thickness less than 1.5 m were considered in our analyses (i.e. undeformed first-year ice; Heil et al. 1996, Worby & Allison 1999). All reported R^2 values are adjusted R^2 to allow comparison

Table 2. Summary statistics for physical sea ice parameters and integrated chl *a* concentrations in sea ice sampled during 3 voyages (where *n* is the number of sampling sites). Reported values are mean \pm standard error. Only cores with ice thickness less than 1.5 m were considered in our analyses (i.e. undeformed first-year pack ice; Heil et al. 1996, Worby & Allison 1999). WWOS: Winter Weddell Outflow Study; SIPEX1, SIPEX2: Sea Ice Physics and Ecosystems Experiments 1 (2007) and 2 (2012)

Parameter	WWOS (n = 19)	SIPEX1 (n = 26)	SIPEX2 (n = 14)
Ice thickness (m)	1.18 \pm 0.05	0.63 \pm 0.06	1.07 \pm 0.06
Snow depth (m)	0.15 \pm 0.04	0.09 \pm 0.02	0.20 \pm 0.04
Chl <i>a</i> (mg m ⁻²)	6.69 \pm 1.29	0.86 \pm 0.19	0.82 \pm 0.18

between models with different numbers of predictors. Given multiple predictors for the EOF approach, we used multiple linear regression with backwards elimination based on Akaike's Information Criterion to select the final models (Burnham & Anderson 2002). To test the applicability of NDI ratios between regions (East Antarctica and the Weddell Sea), we also fitted a relationship for the smaller data set (WWOS) based on the NDI determined for the large data set (SIPEX). All analyses were conducted using the statistical computing software R (R Core Team 2015).

RESULTS

Summary statistics for snow and ice thickness as well as integrated chl *a* are provided in Table 2. Integrated chl *a* concentrations from all 3 expeditions were significantly correlated with ice thickness but showed no correlation with snow depth (Fig. 4; note that integrated chl *a* is presented on a log scale in this figure). Mean integral-normalised transmitted under-ice irradiance spectra for the 3 expeditions and for low (0 to 1 mg m⁻²) and high (1 to 20 mg m⁻²) integrated chl *a* concentrations are presented in Fig. 5. Correlation surfaces of NDIs for integrated chl *a* showed a pronounced switch from negative to positive correlations at 440 nm for both East Antarctica and the Weddell Sea (Fig. 6a,c), consistent with results from Mundy et al. (2007) for the Arctic and with maxima of particulate, ice algal and chl *a* specific absorption (SooHoo et al. 1987, Bricaud et al. 2004, van Leeuwe et al. 2005).

The performances of our 4 alternative approaches are summarised by region in Table 3. All models required log transformation of integrated chl *a* to adjust

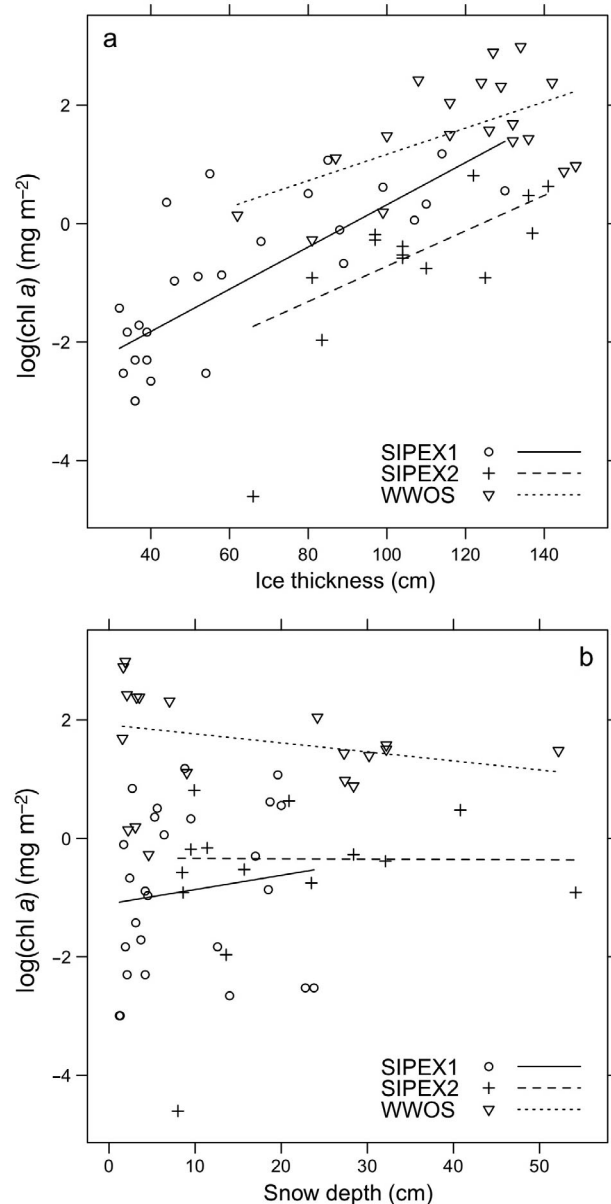


Fig. 4. Relationship between integrated chl *a* and (a) ice thickness and (b) mean snow depth. Two-way ANCOVAs indicate homogeneity of slopes and significant differences in intercepts between voyages for both ice thickness ($F_{2,55} = 19.0$, $p < 0.0001$) and snow depth ($F_{2,55} = 23.1$, $p < 0.0001$). There is also a significant effect of ice thickness on chl *a* ($F_{2,55} = 57.0$, $p < 0.0001$) but no effect of snow depth on chl *a* ($F_{1,55} = 0.19$, $p < 0.68$). SIPEX1, SIPEX2: Sea Ice Physics and Ecosystems Experiments 1 (2007) and 2 (2012); WWOS: Winter Weddell Outflow Study

for heteroscedasticity (i.e. higher variance for higher values) and were significant, with the proportion of variance explained higher than 50%. NDIs had the highest predictive power, and the highest correlations with integrated chl *a* were found for wavelength pairs of 422:418 nm (East Antarctica) and

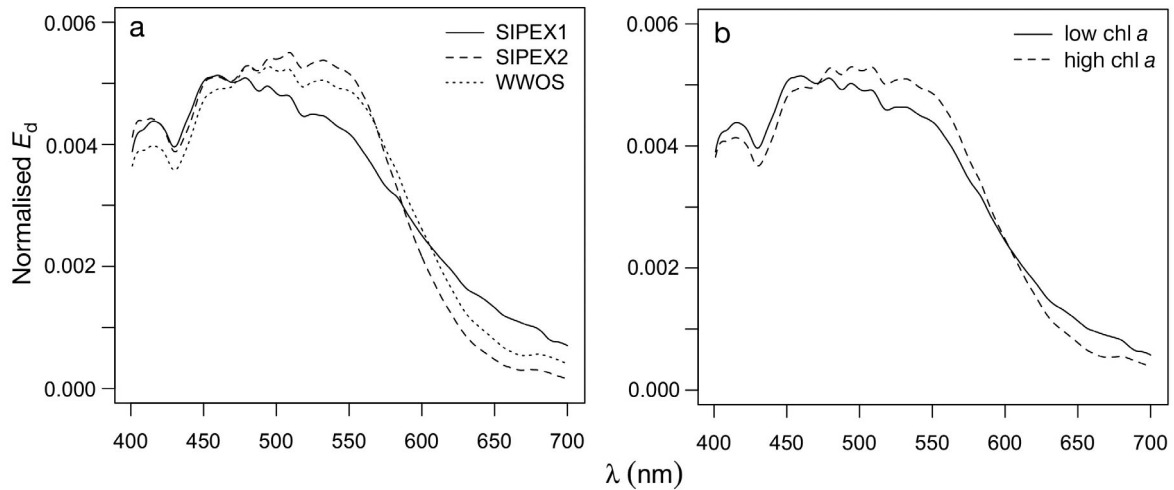


Fig. 5. Mean transmitted under-ice irradiance spectra (normalised to area under curve) for (a) Sea Ice Physics and Ecosystems Experiments 1 (SIPEX1, 2007) and 2 (SIPEX2, 2012) and Winter Weddell Outflow Study (WWOS) and (b) low (0 to 1 mg m^{-2}) and high (1 to 20 mg m^{-2}) integrated chl *a*. E_d : transmitted irradiance; λ : wavelength

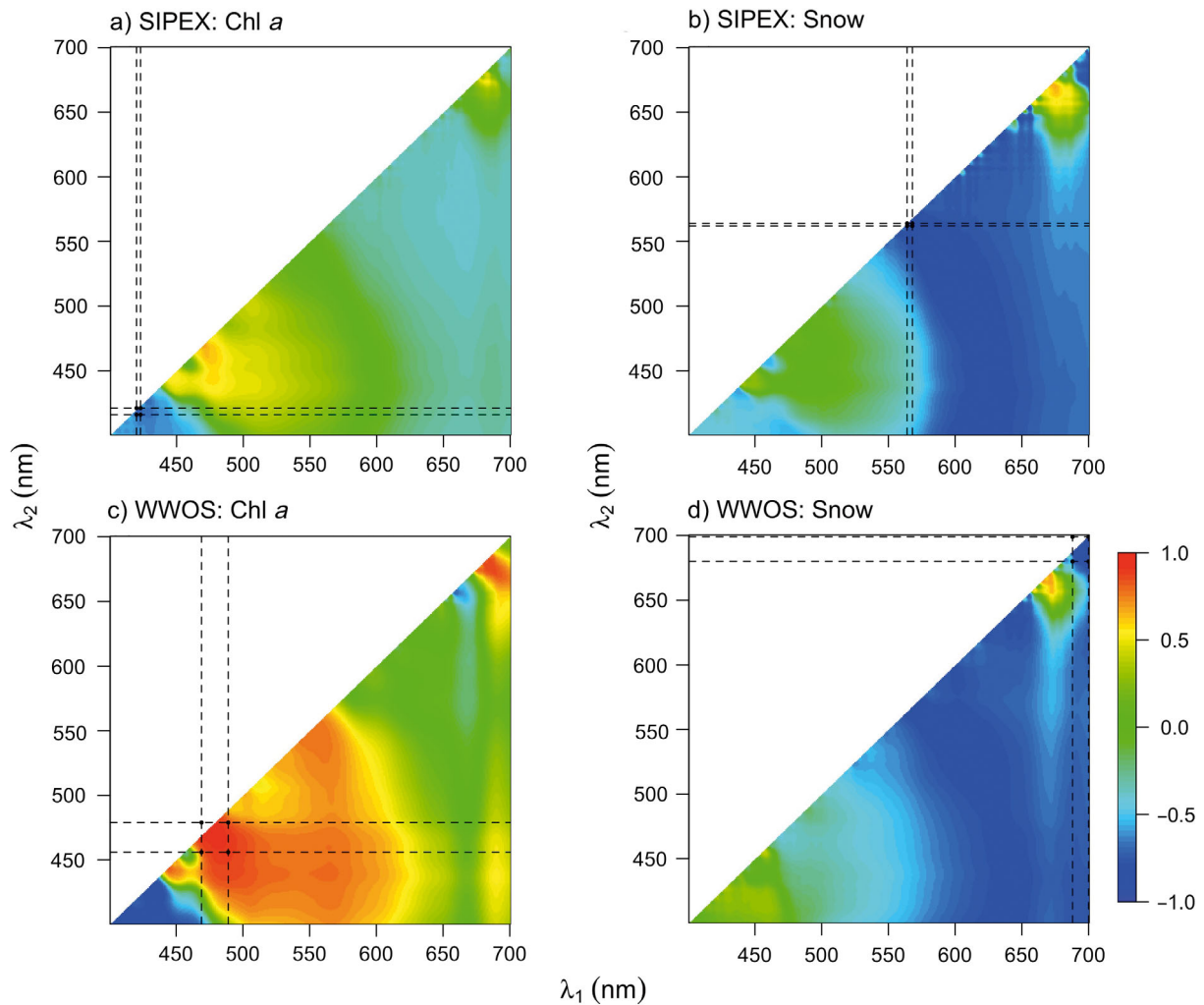


Fig. 6. Correlation surfaces of normalised difference indices for (a,c) integrated chl *a* and (b,d) snow depth for the (a,b) Sea Ice Physics and Ecosystems Experiments (SIPEX1 and SIPEX2, East Antarctica) and (c,d) Winter Weddell Outflow Study (WWOS, Weddell Sea) data. Dashed lines identify regions of these surfaces with the highest absolute correlation values. λ_1 , λ_2 : wavelength pairs (see Eq. 1)

Table 3. Comparison of algorithms for estimating integrated chl *a*. For the empirical orthogonal function (EOF) methods, S1 to S4 are the scores of the first 4 EOFs. SIPEX1, SIPEX2: Sea Ice Physics and Ecosystems Experiments 1 (2007) and 2 (2012); NDI: normalised difference index; E_d : transmitted irradiance; sba: scaled band area; WWOS: Winter Weddell Outflow Study

Region	Method	Observed relationship	R ²	p
East Antarctica (SIPEX1, SIPEX2)	NDI	$\ln(\text{chl } a) = -4.27 - 351 \times \text{NDI}(422:418)$	0.64	<0.0001
	$E_d(555)/E_d(443)$	$\ln(\text{chl } a) = -1103 + 1948 \times E_d(555)/E_d(443) - 859 \times [E_d(555)/E_d(443)]^2$	0.56	<0.0001
	Scaled band area	$\ln(\text{chl } a) = -16.36 + 9.52 \times \text{sba} - 1.34 \times \text{sba}^2$	0.64	<0.0001
	EOFs	$\ln(\text{chl } a) = 0.36 + 6.41 \times S1 - 143.5 \times S2 - 20970 \times S2^2 + 393.3 \times S3 - 512.6 \times S4$	0.52	<0.0001
Weddell Sea (WWOS)	NDI	$\ln(\text{chl } a) = 0.39 + 31.7 \times \text{NDI}(479:468)$	0.79	<0.0001
	$E_d(555)/E_d(443)$	$\ln(\text{chl } a) = -33.9 + 31.0 \times E_d(555)/E_d(443)$	0.67	0.0001
	Scaled band area	$\ln(\text{chl } a) = -2.40 + 1.64 \times \text{sba} - 0.13 \times \text{sba}^2$	0.60	0.0003
	EOFs	$\ln(\text{chl } a) = 1.55 + 43.0 \times S1 + 112.5 \times S2 - 243.7 \times S3$	0.67	0.0001

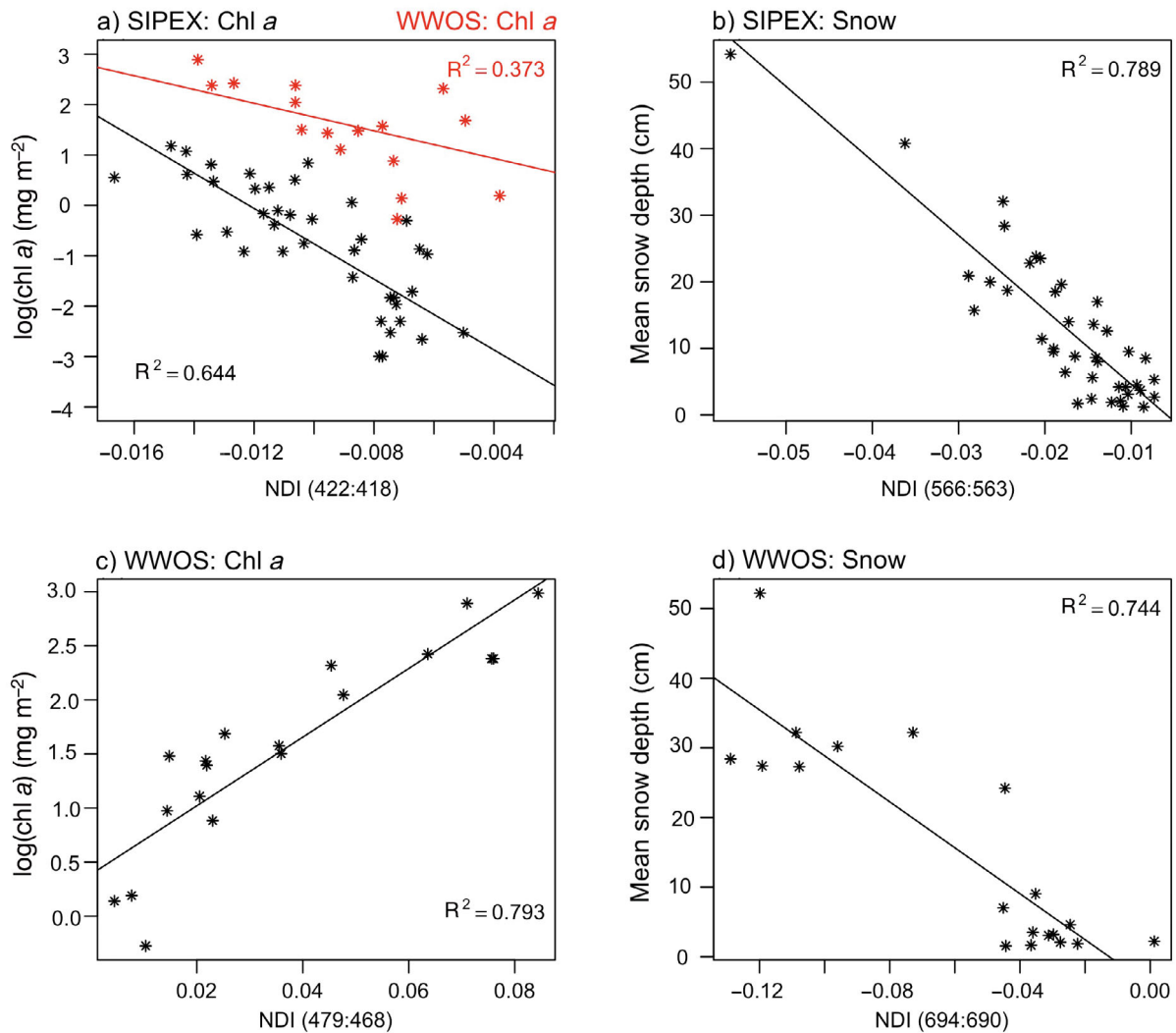


Fig. 7. Relationships between observed normalised difference indices (NDIs) and (a,c) integrated chl *a* and (b,d) mean snow depth for the (a,b) Sea Ice Physics and Ecosystems Experiments (SIPEX1 and SIPEX2, East Antarctic) and (c,d) Winter Weddell Outflow Study (WWOS, Weddell Sea). Wavelength pairs (nm) determined from examination of correlation surfaces (Fig. 6) are indicated in x-axis labels. Equations and significance of fitted relationships are provided in Table 3. The fitted relationship for WWOS integrated chl *a* based on the NDI (422:418 nm) derived from SIPEX is shown in red in (a), $p = 0.003$

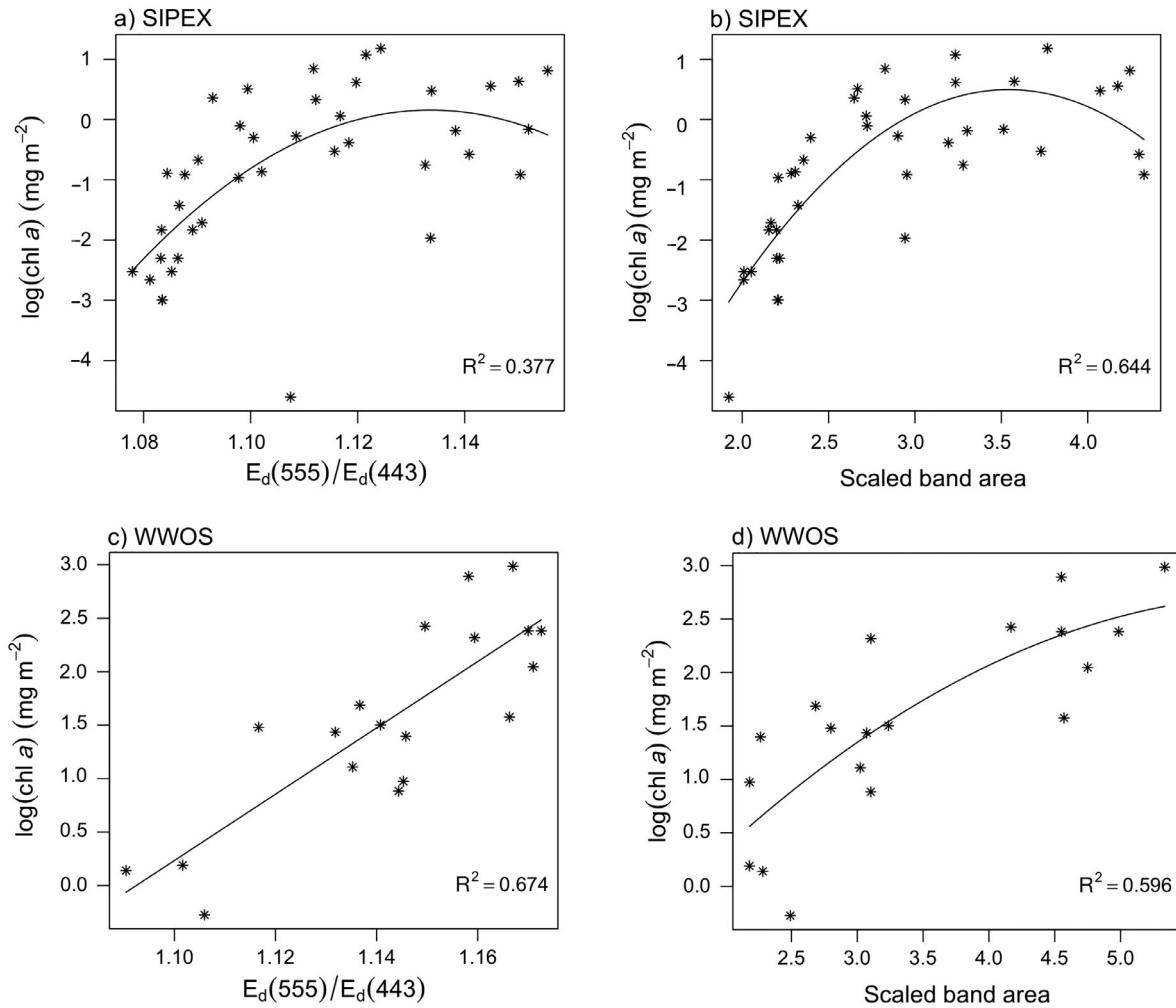


Fig. 8. Relationships between observed integrated chl *a* and (a,c) transmitted irradiance ratio $E_d(555)/E_d(443)$ and (b,d) scaled band area for the (a,b) Sea Ice Physics and Ecosystems Experiments (SIPEX1 and SIPEX2, East Antarctica) and (c,d) Winter Weddell Outflow Study (WWOS, Weddell Sea). The removal of a single outlier in (a) gave an R^2 value of 0.56 as reported in Table 3. All equations and significance of fitted relationships are provided in Table 3

479:468 nm (Weddell Sea). Wavelength pairs that gave high correlations with snow depth (566:563 nm, $R^2 = 0.79$ for East Antarctica and 694:690 nm, $R^2 = 0.74$ for the Weddell Sea; see Fig. 7) were non-overlapping with NDI wavelengths for chl *a*. We note that single outlying points in SIPEX NDI regressions for chl *a* (Fig. 7a) and snow (Fig. 7b) are 2 different sampling sites. Removal of these sites from respective analyses still produced significant regressions with similar R^2 values. The model for WWOS integrated chl *a* based on the NDI ratio (422:418 nm) determined from SIPEX explained 37% of the variability in observed chl *a* for the Weddell Sea. The fitted relationship is shown in red in Fig. 7a.

Models relating chl *a* to scaled band area required linear and quadratic terms for both regions (Table 3,

Fig. 8) and explained on the order of 60% of the variation in chl *a*. The $E_d(555)/E_d(443)$ model for East Antarctica also required a quadratic term and explained 38% of the variation in chl *a* (56% when a single outlier with low chl *a* was removed). The worst-performing algorithm by region combination was the EOF analysis for East Antarctica. EOF analysis and model fitting for the 2 separate voyages in East Antarctica gave significant fits, with R^2 values of 0.86 and 0.82 for SIPEX1 and SIPEX2, respectively. However, these models had very different formulations, with Modes 1 to 4 as predictors for SIPEX1 and only the second and fourth modes as predictors for SIPEX2.

The modes of oscillation in EOF loadings (Figs. 9 & 10) can be interpreted as signatures of changes in

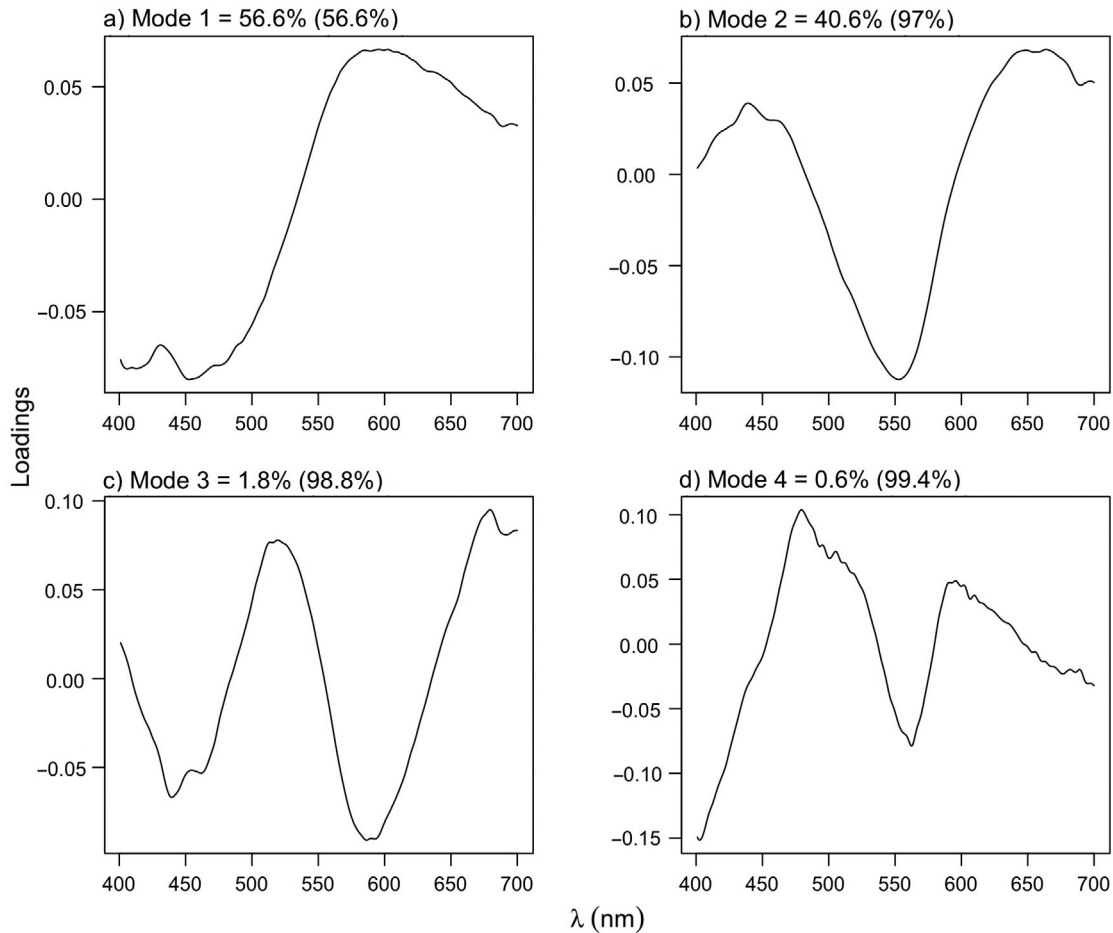


Fig. 9. Loadings, percent variance and cumulative proportion of variance represented (in parentheses) for Modes 1 to 4 of empirical orthogonal function analysis of transmitted spectra for the Sea Ice Physics and Ecosystems Experiments (SIPEX1 and SIPEX2, East Antarctica). λ : wavelength

the optical properties of absorbing constituents (as demonstrated by Craig et al. 2012 and Taylor et al. 2013). For the Weddell Sea, Mode 1 includes the chl *a* fluorescence emission peak at ~683 nm noted by Craig et al. (2012) as a signature of changes in chl *a* (Fig. 10a). Mode 2 for this region (Fig. 10b) resembles Mode 1 but with a change in the sign of the loading at around 600 nm, which may be associated with differences in the degree of scattering by snow. Spectral inflections in Modes 3 and 4 may be attributable to variability in phytoplankton pigment composition, although these modes capture very small proportions of the total variability in spectral shape (0.7 and 0.6%), and only Modes 1 to 3 were included in the fitted multiple regression model for chl *a* (Table 3). For East Antarctica, Modes 2 and 4 were similar in shape (Fig. 9b,d) and exhibited relatively strong negative loadings at ~550 nm. As for the Weddell Sea, inflections in Mode 3 for this region (Fig. 10c) may be associated with variable pigment compositions. All 4

modes were included in the final fitted model for East Antarctica (Table 3).

DISCUSSION

Given the importance of sea ice habitats in Antarctic marine ecosystems (Arrigo 2014) and the potential but currently uncertain contribution of ice algae to overall Southern Ocean primary productivity (Lizotte 2001, Saenz & Arrigo 2014), there is a strong need for improved estimates of ice algal biomass distribution at meso- to regional scales. In this study, we evaluated alternative approaches to estimating chl *a* from bio-optical measurements under Antarctic sea ice in 2 regions: the Weddell Sea and East Antarctica. Our results indicate that NDIs provide the most robust predictions of integrated chl *a*, both at the regional scale and as a general method that is applicable across multiple regions.

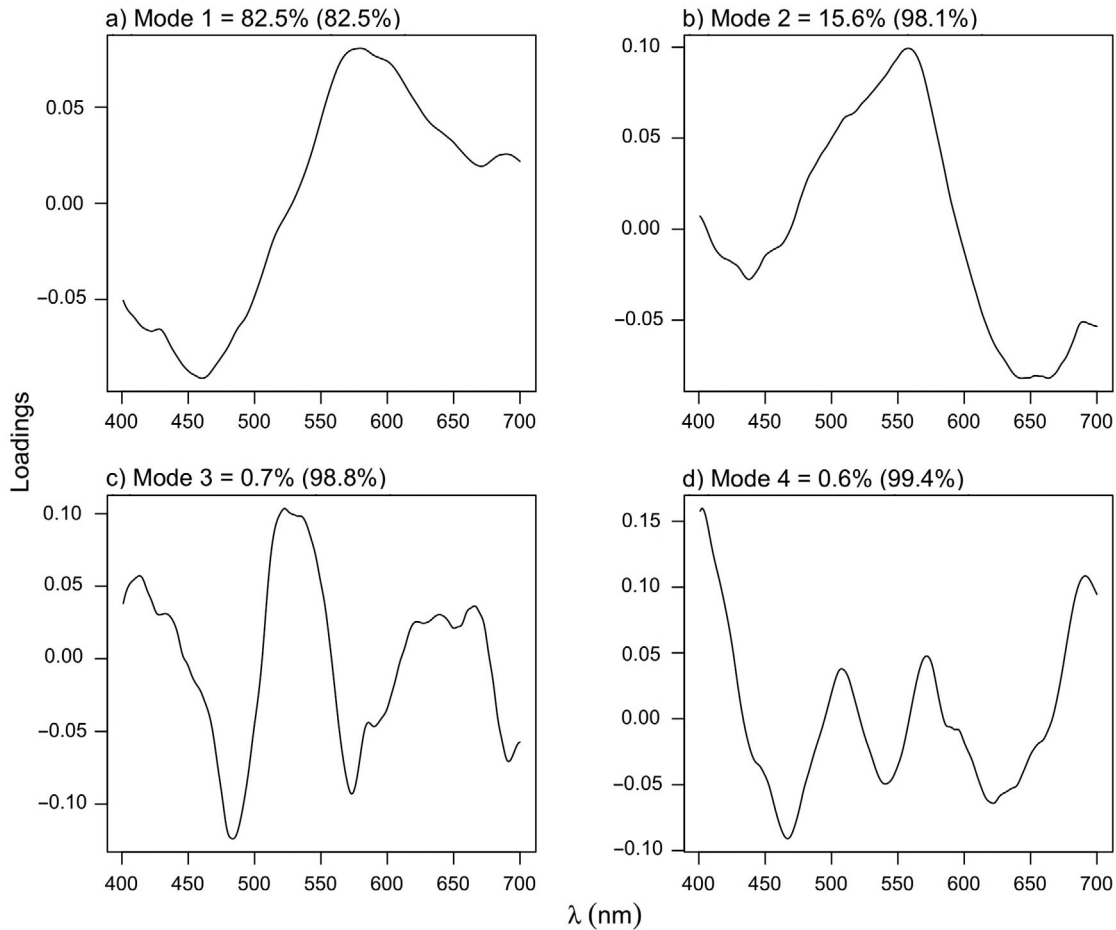


Fig. 10. Loadings, percent variance and cumulative proportion of variance represented (in parentheses) for Modes 1 to 4 of empirical orthogonal function analysis of transmitted spectra for the Winter Weddell Outflow Study (WWOS, Weddell Sea). λ : wavelength

We observed a log-linear relationship between integrated chl *a* and ice thickness (for ice cores less than 1.5 m length), with equivalent slopes between data sets (Fig. 4), but no significant relationship between chl *a* and snow depth. Strong inverse relationships between snow cover thickness and ice algal biomass have been reported from both Antarctic and Arctic fast ice habitats in previous studies (e.g. Palmisano et al. 1987, Mundy et al. 2007). We interpret the lack of correlation between snow cover and ice algal biomass in our pack ice dataset to be a result of high snow cover dynamics in the Weddell Sea and East Antarctica, i.e. pack ice snow cover is affected by lateral transport, causing strong redistribution resulting in high temporal variability in winter (Massom et al. 1997, 1998) and most likely also in early spring.

Integrated chl *a* values reported in this study (~ 1 to 7 mg m^{-2} ; Table 2) are much lower than those reported by Campbell et al. (2015) and Mundy et al. (2007) for the Arctic (0 to 100 mg m^{-2}) and at the lower end of the scale compared with seasonal

means from historical circumpolar Antarctic ice core data (0 to 13 mg m^{-2} ; Meiners et al. 2012). Low integrated biomass will result in less absorption of transmitted irradiance spectra, thereby influencing the signal-to-noise ratio (e.g. by enhancing the influence of snow cover as demonstrated by Mundy et al. 2007 for the Arctic) and reducing the sensitivity of the derived algorithms. Applying the bio-optical approaches of this study to high sea ice algal biomass habitats, e.g. coastal Antarctic fast ice, may provide even more robust relationships between ice algal chl *a* concentrations and under-ice irradiance spectra.

Algorithms for estimating chl *a*

The best-performing algorithm evaluated was the NDI approach, with NDI models explaining ~ 60 to 80% of observed variation in chl *a* (Table 3). When the NDI ratio identified for the larger data set (SIPEX) was used to fit an algorithm for the smaller

Table 4. Comparison of normalised difference index (NDI) wavelength ratios and corresponding algorithms for the present study, Campbell et al. (2014) and Mundy et al. (2007) for the Arctic. Note that Mundy et al. used chl *a* as the predictor and the NDI ratio as the response; equations have been transformed here to match the form used in our study, but R^2 values are not comparable. SIPEX: Sea Ice Physics and Ecosystems Experiment; WWOS: Winter Weddell Outflow Study

Study/location	NDI ratio	Algorithm	R^2
Present study/East Antarctica (SIPEX)	NDI(422:418)	$\ln(\text{chl } a) = -4.27 - 351 \times \text{NDI}(422:418)$	0.64
Present study/Weddell Sea (WWOS)	NDI(479:468)	$\ln(\text{chl } a) = -0.39 - 31.7 \times \text{NDI}(479:468)$	0.79
Campbell et al. (2014)	NDI(478:490)	$\text{chl } a = 15.2 - 497 \times \text{NDI}(478:490)$	0.81
Mundy et al. (2007)	NDI(415:400)	$\text{chl } a = 80.2 - 588 \times \text{NDI}(415:400)$	0.81 ^a
	NDI(485:472)	$\text{chl } a = -8.3 + 1000 \times \text{NDI}(485:472)$	0.89 ^a

^aNDI is the predictor, i.e. $\text{NDI}(415:400) = -0.0017\text{chl } a + 0.1364$ and $\text{NDI}(485:472) = 0.001\text{chl } a + 0.0083$

data set (WWOS), the fit was significant ($p = 0.003$) and explained approximately 40% of the variability in chl *a* (Fig. 7a). Mundy et al. (2007) identified the wavelength pairs 415:400 and 485:472 nm as the best predictors of chl *a* in Arctic fast ice, with fitted models explaining 81 and 89% of the variance. Our analyses identified similar NDI ratios of 422:418 and 479:468 nm as the best predictors for East Antarctica and the Weddell Sea, respectively (see Table 4 for a comparison of wavelength ratios and algorithms). Again using NDI wavelength ratios from a larger data set (Mundy et al. 2007, $n = 47$) to fit algorithms for smaller data sets (present study), we found that Mundy et al.'s NDI ratios performed well for the Weddell Sea (explaining 70 and 79% of the variance) but not as well for East Antarctica (explaining only 45 and 32% of the variance). This difference may be associated with lower integrated chl *a* concentrations for East Antarctica (Table 2) as compared with the Weddell Sea and Mundy et al.'s measurements. We suggest that NDIs derived from correlation surfaces perform best among the algorithms we evaluated because they effectively target particular pigments rather than integrating signals across multiple pigments (as summarised in Table 1).

The $E_d(555)/E_d(443)$ ratio, scaled band area and EOFs provided reasonable predictions of chl *a* at the regional level. Of these 3 methods, EOFs have the advantage of interpretable patterns in EOF loadings, although this would ideally be coupled with measurements of phytoplankton, particulate and coloured dissolved organic matter (CDOM) absorption coefficients for full interpretation (as in Craig et al. 2012). The performance of the scaled band area method (R^2 values of 0.64 and 0.60 for East Antarctica and the Weddell Sea, respectively) was surprisingly good given that previous applications of this approach (Carrere et al. 2004) were for reflectance spectra. The $E_d(555)/E_d(443)$ ratio also demonstrated reasonable

predictive power, particularly for the smaller Weddell Sea dataset ($R^2 = 0.67$). We note that Fritsen et al. (2011) successfully used a slight variant of this ratio, $E_d(555)/E_d(442)$, for assessing changes in sea ice algal biomass in the Bellingshausen Sea; these wavelengths correspond with spectral bands tested in the Fritsen et al. study.

What can transmitted spectra tell us about Antarctic pack ice habitats?

In ice-covered areas, the transmission of solar radiation through sea ice and into the upper ocean is determined by spatial and temporal variations in incident irradiance, ice concentration, snow and ice thickness, and the optical properties of the snow and sea ice. Snow cover strongly affects overall transmission of solar radiation because snow extinction coefficients are more than an order of magnitude larger than those of bare sea ice, and snow albedos can be almost twice as large as those of ice (Grenfell & Maykut 1977). Other important factors controlling transmission of light through sea ice are its crystalline structure, gas and brine inclusions (which act as scatterers of light; Light et al. 2004) and impurities, for example CDOM (which acts as an absorber in the 280 to 450 nm range; Belzile et al. 2000). Variability in these features between locations may influence our ability to develop cross-site relationships between transmitted spectra and chl *a*, although the use of particular wavelength ratios is designed to mitigate the effects of such confounding factors.

In contrast to Arctic pack and in particular fast ice, Antarctic pack ice harbours high biomass in surface, interior and bottom communities (Thomas & Dieckmann 2010, Meiners et al. 2012). Ice algal communities in different depth horizons of the sea ice are exposed to diverse light levels where they use differ-

ent acclimation strategies that likely strongly affect the transmitted light versus chl *a* relationship. For example, shade acclimation of ice algae can lead to increases in chl *a* per cell and increased production of accessory pigments (SooHoo et al. 1987, Alou-Font et al. 2013) that can increase the package effect (i.e. decreased absorption efficiency per chl *a*; Falkowski & Raven 2007). Alternatively, high light acclimation can lead to the opposite effect of a decreased package effect due to less chl *a* per cell. In the context of our study, stations with similar integrated chl *a* concentrations may therefore have variable optical properties due to different integrated community composition when compared among each other and to Arctic sea ice.

Forward scattering of light in sea ice refers to the process by which surface irradiance is funnelled and focussed towards a smaller area when transmitted from the surface to the bottom layers of sea ice (due to brine and salt inclusions; Light et al. 2004). Surface algal communities in proximity to our collected ice cores may therefore have influenced under-ice irradiance and might not have been properly quantified by our single-core *in situ* sampling strategy. To quantify this effect, we suggest the sampling of several ice cores at the radiometer site during future studies and also the alternative use of radiance sensors with smaller footprints than cosine sensors. Overall, the comparison of radiometer data and ice core data—and the associated development of algorithms—are impacted by the representativeness of ice core chl *a* data for the entire footprint sampled by the respective radiometer.

Under-ice phytoplankton biomass may also have affected our measurements. Under-ice phytoplankton biomass was very low during all voyages (<0.2 mg chl *a* m^{-3}) and therefore accounted for a maximum of 3.75% of the integrated algal biomass above the sensor (Table 2). Future use of moored optical sensors and instrumented vehicles needs to consider potential effects of under-ice algal biomass, which can be high and can also accumulate in aggregates and strands, in particular under Arctic sea ice (Mundy et al. 2011, Arrigo et al. 2012, Boetius et al. 2013), which will affect the applicability of the described optical methods.

CONCLUSIONS

Our study is the first to compare algorithms for estimating Antarctic sea ice algal biomass using under-ice irradiance spectra at a regional scale. Our results

demonstrate the strong potential to estimate Antarctic ice algal biomass from transmitted under-ice irradiance spectra, in particular using NDIs. Given some degree of regional dependency for our best-performing method (NDIs), our results suggest that this method may be more appropriate for time series measurements using moored sensors than multi-regional mapping efforts (Nicolaus et al. 2013, Campbell et al. 2015). However, explanatory power might be improved through consideration of additional predictive variables (e.g. the vertical distribution of ice algal biomass, the biomass of under-ice phytoplankton or the concentration of non-algal particles).

Nicolaus et al. (2013) found that the seasonal evolution of transmitted solar radiation through landfast sea ice near Barrow, Alaska, USA, exceeded the spatial variability. All 3 of the voyages considered here were carried out early in the season during spring algal biomass buildup. Further work is needed to study interrelationships between ice algal biomass, sea ice particulate and ice algal absorption, and under-ice irradiance throughout the peak and decline of ice algal blooms. For example, senescence of algae is generally accompanied by an increase in chl *a* degradation products such as pheophytin *a* (Alou-Font et al. 2013), which shows increased absorption maxima at shorter wavelengths (410 nm). While the absorption of non-algal particles and coloured dissolved inorganic matter also increases with shorter wavelengths, it might be possible to not only monitor ice algal biomass development but also estimate ice algal health through analysis of under-ice spectra over time. Further research would also usefully link this work to our understanding of changes in the Antarctic energy budget, as has been done for the Arctic (e.g. Nicolaus et al. 2012).

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LITERATURE CITED

Ackley SF, Lewis MJ, Fritsen CH, Xie H (2008) Internal melting in Antarctic sea ice: development of 'gap layers'. *Geophys Res Lett* 35:L11503, doi:10.1029/2008GL033644

- Alou-Font E, Mundy CJ, Roy S, Gosselin M, Agustí S (2013) Snow cover affects ice algal pigment composition in the coastal Arctic Ocean during spring. *Mar Ecol Prog Ser* 474:89–104
- Arar EJ, Collins GB (1997) Method 445.0: *in vitro* determination of chlorophyll *a* and pheophytin *a* in marine and freshwater algae by fluorescence. In: *Methods for the determination of chemical substances in marine and estuarine environmental matrices*, 2nd edn. EPA/600/R-97/072. National Exposure Research Laboratory, Office of Research and Development, USEPA, Cincinnati, OH, p 445.0-1–445.0.22
- Arrigo KR (2014) Sea ice ecosystems. *Annu Rev Mar Sci* 6: 439–467
- Arrigo KR, Robinson DH, Sullivan CW (1993) A high resolution study of the platelet ice ecosystem in McMurdo Sound, Antarctica: photosynthetic and bio-optical characteristics of a dense microalgal bloom. *Mar Ecol Prog Ser* 98:173–185
- Arrigo KR, Mock T, Lizotte MP (2010) Primary producers and sea ice. In: Thomas DN, Dieckmann GS (eds) *Sea ice*. Wiley-Blackwell, Chichester, p 283–326
- Arrigo KR, Perovich DK, Pickart RS, Brown ZW and others (2012) Massive phytoplankton blooms under Arctic sea ice. *Science* 336:1408
- Beeler SooHoo J, Palmisano AC, Kottmeier ST, Lizotte MP, SooHoo SL, Sullivan CW (1987) Spectral light absorption and quantum yield of photosynthesis in sea ice microalgae and a bloom of *Phaeocystis pouchetii* from McMurdo Sound, Antarctica. *Mar Ecol Prog Ser* 39:175–189
- Belzile C, Johannessen SC, Gosselin M, Demers S, Miller WL (2000) Ultraviolet attenuation by dissolved and particulate constituents of first-year ice during late spring in an Arctic polynya. *Limnol Oceanogr* 45:1265–1273
- Boetius A, Albrecht S, Bakker K, Bienhold C and others (2013) Export of algal biomass from the melting Arctic sea ice. *Science* 339:1430–1432
- Bricaud A, Claustre H, Ras J, Oubelkheir K (2004) Natural variability of phytoplanktonic absorption in oceanic waters: influence of the size structure of algal populations. *J Geophys Res* 109:C11010, doi:10.1029/2004JC002419
- Burnham KP, Anderson DR (2002) *Model selection and multimodel inference: a practical information-theoretic approach*. Springer, New York, NY
- Campbell K, Mundy CJ, Barber D, Gosselin M (2015) Response of remotely estimated ice algae biomass to the environmental conditions during spring melt. *J Mar Syst* 147:76–84
- Carrère V, Spilmont N, Davoult D (2004) Comparison of simple techniques for estimating chlorophyll *a* concentration in the intertidal zone using high spectral-resolution field-spectrometer data. *Mar Ecol Prog Ser* 274:31–40
- Craig SE, Jones CT, Li WKW, Lazin G, Horne E, Caverhill C, Cullen JJ (2012) Deriving optical metrics of coastal phytoplankton biomass from ocean colour. *Remote Sens Environ* 119:72–83
- Falkowski PG, Raven JA (2007) *Aquatic photosynthesis*. Princeton University Press, Princeton, NJ
- Flores H, van Franeker JA, Siegel V, Haraldsson M and others (2012) The association of Antarctic krill *Euphausia superba* with the under-ice habitat. *PLoS ONE* 7:e31775
- Fritsen CH, Wirthlin ED, Momborg DK, Lewis MJ, Ackley SF (2011) Bio-optical properties of Antarctic pack ice in the early austral spring. *Deep-Sea Res II* 58:1052–1061
- Grenfell TC, Maykut GA (1977) The optical properties of ice and snow in the Arctic basin. *J Glaciol* 18:445–463
- Hawes I, Lund-Hansen LC, Sorrell BK, Nielsen MH, Borzak R, Buss I (2012) Photobiology of sea ice algae during initial spring growth in Kangerlussuaq, West Greenland: insights from imaging variable chlorophyll fluorescence of ice cores. *Photosynth Res* 112:103–115
- Heil P, Allison I, Lytle VI (1996) Seasonal and interannual variations of the oceanic heat flux under a landfast Antarctic sea ice cover. *J Geophys Res Oceans* 101: 25741–25752
- Holm-Hansen O, Lorenzen CJ, Holmes RW, Strickland JDH (1965) Fluorometric determination of chlorophyll. *ICES J Mar Sci* 30:3–15
- Honda MC, Sasaoka K, Kawakami H, Matsumoto K, Watanabe S, Dickey T (2009) Application of underwater optical data to estimation of primary productivity. *Deep-Sea Res I* 56:2281–2292
- Legendre L, Gosselin M (1991) In situ spectroradiometric estimation of microalgal biomass in first-year sea ice. *Polar Biol* 11:113–115
- Lemke P (ed) (2009) *The expedition of the research vessel 'Polarstern' to the Antarctic in 2006 (ANT-XXIII/7)*. Reports on Polar and Marine Research, Alfred Wegener Institute for Polar and Marine Research, Bremerhaven
- Light B, Maykut GA, Grenfell TC (2004) A temperature-dependent, structural-optical model of first-year sea ice. *J Geophys Res* 109:C06013, doi:10.1029/2003JC002164
- Lizotte MP (2001) The contributions of sea ice algae to Antarctic marine primary production. *Am Zool* 41:57–73
- Massom RA, Drinkwater MR, Haas C (1997) Winter snow cover on sea ice in the Weddell Sea. *J Geophys Res Oceans* 102:1101–1117
- Massom RA, Lytle VI, Worby AP, Allison I (1998) Winter snow cover variability on East Antarctic sea ice. *J Geophys Res Oceans* 103:24837–24855
- McDonald S, Koullis T, Ehn J, Campbell K, Gosselin M, Mundy CJ (2015) A functional regression model for predicting optical depth and estimating attenuation coefficients in sea-ice covers near Resolute Passage, Canada. *Ann Glaciol* 56:147–154
- Meiners KM, Vancoppenolle M, Thanassekos S, Dieckmann GS and others (2012) Chlorophyll *a* in Antarctic sea ice from historical ice core data. *Geophys Res Lett* 39: L21602, doi:10.1029/2012GL053478
- Mundy CJ, Ehn JK, Barber DG, Michel C (2007) Influence of snow cover and algae on the spectral dependence of transmitted irradiance through Arctic landfast first-year sea ice. *J Geophys Res* 112:1–10
- Mundy CJ, Gosselin M, Ehn JK, Belzile C and others (2011) Characteristics of two distinct high-light acclimated microbial communities during advanced stages of sea ice melt. *Polar Biol* 34:1869–1886
- Nicolaus M, Katlein C, Maslanik J, Hendricks S (2012) Changes in Arctic sea ice result in increasing light transmittance and absorption. *Geophys Res Lett* 39:L24501, doi:10.1029/2012GL053738
- Nicolaus M, Petrich C, Hudson SR, Granskog MA (2013) Variability of light transmission through Arctic landfast sea ice during spring. *Cryosphere* 7:977–986
- Palmisano AC, Beeler SooHoo J, Moe RL, Sullivan CW (1987) *Sea ice microbial communities*. VII. Changes in under-ice spectral irradiance during the development of Antarctic sea ice microalgal communities. *Mar Ecol Prog Ser* 35:165–173

- Parkinson CL, Cavalieri DJ (2012) Antarctic sea ice variability and trends, 1979–2010. *Cryosphere* 6:871–880
- Perovich DK, Cota GF, Maykut GA, Grenfell TC (1993) Bio-optical observations of first-year Arctic sea ice. *Geophys Res Lett* 20:1059–1062
- R Core Team (2015) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna. Available at www.R-project.org/
- Saenz BT, Arrigo KR (2014) Annual primary production in Antarctic sea ice during 2005–2006 from a sea ice state estimate. *J Geophys Res Oceans* 119:3645–3678
- Taylor BB, Taylor MH, Dinter T, Bracher A (2013) Estimation of relative phycoerythrin concentrations from hyperspectral underwater radiance measurements—a statistical approach. *J Geophys Res Oceans* 118:2948–2960
- Thomas DN, Dieckmann GS (eds) (2010) *Sea ice*, 2nd edn. Wiley-Blackwell, Chichester
- van Leeuwe MA, van Sikkelerus B, Gieskes WWC, Stefels J (2005) Taxon-specific differences in photoacclimation to fluctuating irradiance in an Antarctic diatom and a green flagellate. *Mar Ecol Prog Ser* 288:9–19
- Vancoppenolle M, Goosse H, de Montety A, Fichetef T, Tremblay B, Tison JL (2010) Modeling brine and nutrient dynamics in Antarctic sea ice: the case of dissolved silica. *J Geophys Res Oceans* 115:C02005, doi:10.1029/2009JC005369
- Vancoppenolle M, Meiners KM, Michel C, Bopp L and others (2013) Role of sea ice in global biogeochemical cycles: emerging views and challenges. *Quat Sci Rev* 79:207–230
- Worby AP, Allison I (1999) A technique for making ship-based observations of Antarctic sea ice thickness and characteristics. Part I: observational technique and results. Res Rep No. 14, Antarctic Cooperative Research Centre, Hobart
- Worby AP, Steer A, Lieser JL, Heil P and others (2011) Regional-scale sea-ice and snow thickness distributions from in situ and satellite measurements over East Antarctica during SIPEX 2007. *Deep-Sea Res II* 58:1125–1136

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