



The interplay between animal location accuracy and the decorrelation length scale of environmental variables when investigating environmental selection in marine organisms

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ABSTRACT: Many large pelagic organisms appear to select specific oceanic conditions, probably due to physiological, energetic, reproductive, or other life history needs. However, prior to characterizing these dynamics and determining their underlying drivers, the selection itself must be reliably identified. Our ability to do so depends on the quality of animal locations and on the heterogeneity of the environmental conditions driving selection. To draw meaningful conclusions about environmental selection of large organisms and therefore about their basic ecology, distribution, and ultimately their potential exploitation and conservation, limits of selection detectability must be established. Here, we investigated how animal location accuracy and environmental variable decorrelation length scales impact the ability to detect environmental selection by marine organisms. We created synthetic tracks cuing on environmental variables potentially relating to life history demands, like sea surface temperature, chlorophyll *a* concentration, and Lagrangian coherent structures. By artificially imposing different animal selection strengths and location accuracies, we assessed how well environmental selection can be detected statistically in different cases. We found medium or strong selection to be reliably detected, even with relatively small samples and large position uncertainty, while weak selection presented similarly to no selection, especially for large sample sizes and position uncertainties. We therefore recommend using a selection strength threshold, which can significantly reduce the number of false positives while only increasing the risk of false negatives in cases of very weak selection, which are also less meaningful ecologically. We provide criteria to use when assessing confidence in environmental selection results for real marine organisms.

KEY WORDS: Movement ecology · Habitat selection · Environmental selection · Biotelemetry · Decorrelation length scale

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1. INTRODUCTION

Marine ecosystems provide humanity with a number of important ecosystem services, including tourism and furnishing food for a growing portion of the world's population (Peterson & Lubchenco 1997). However, marine ecosystems (and consequently the

services they provide) are increasingly threatened by pollution, overexploitation, habitat destruction, and climate change (Dulvy et al. 2003, Lotze et al. 2006, Halpern et al. 2008). Preserving marine ecosystems in the face of continued and increasing human exploitation requires careful spatial planning and management. Effective management is only possible if research

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chers and management agencies are able to observe and characterize the spatial ecology of marine species and ecosystems. Importantly, assessing the spatial ecology of oceanic species has historically been very difficult due to the sheer scale of their habitats (i.e. the open ocean), their highly mobile nature, the dynamic nature of oceanic ecosystems, and, perhaps most importantly, the logistical and technological difficulties associated with actually studying these animals in the wild (Block et al. 2002).

Understanding how mobile marine organisms react to various oceanographic features is critical for informing management of these species and ecosystems. Marine organisms may favor and select more or less strongly for specific environmental conditions, such as specific ranges of sea surface temperature (SST) or chlorophyll *a* (chl *a*) concentration that can be tracked remotely from satellites (Abrahms et al. 2019, Lee et al. 2021, Pinti et al. 2022). By associating the distribution of marine species with particular environmental conditions, we can characterize environmental selection in these animals and predict environmental mismatches that may arise because of climate change (Pörtner & Knust 2007, Møller et al. 2008). The first step to predicting the adaptive capacity of an ecosystem is to identify environmental selection (Tew Kai et al. 2009, Oliver et al. 2019, Hazen et al. 2021, Fahlbusch et al. 2022) and to separate the selection signal from observation errors. These errors may arise from position uncertainty for animal locations (Braun et al. 2023) or from a mismatch of the scales at which animals react to environmental variables and those at which they are observed (Scales et al. 2017).

To estimate animal locations, a wide array of electronic tag technologies are available, each with its own specificities and with different spatio-temporal resolutions and accuracies (Hussey et al. 2015). For example, acoustic telemetry records precisely when an animal carrying an acoustic tag is within the range of an acoustic receiver, but does not provide any information when animals are outside those detection ranges (Matley et al. 2022). Pop-up satellite archival tags (PSATs) rely on light levels to compute time of dawn and dusk, and day duration (sometimes coupled with SST or other measurements) to estimate the position of individuals, but that means that position uncertainties routinely reach hundreds of kilometers (Gunn et al. 1994, Sibert et al. 2003). Positions obtained from ARGOS tags are generally much more precise (~100s of meters to kilometers), although their uncertainty can also sometimes reach 10s to 100s of kilometers. Additionally, ARGOS tags require the animal to stay at the surface for some period of time so that

multiple ARGOS satellites can triangulate the tag position (Douglas et al. 2012). The development of Fastloc GPS tags has facilitated the collection of much more precise location data with uncertainties only up to a few meters, even for animals that only surface briefly (Dujon et al. 2014, Thomson et al. 2017). While these technologies all provide similar types of data, the large range of uncertainties associated with their respective location estimates means that they cannot be used interchangeably and that the ecological question that is being investigated should dictate the kind of biologging technology used. As PSATs are often the only possibility for studies of pelagic organisms that do not surface, large uncertainties are a particular challenge for assessing habitat selection in marine organisms. Using tracks with very large location uncertainty may yield incorrect results or blur selection signals.

Environmental conditions also vary in space and time, and their measurements come with their own uncertainties. Decorrelation length scales (DCLSs) estimate the spatial distance over which a variable (e.g. temperature) remains correlated. Formally, the DCLS is the distance at which the correlation between the time series of a variable (or its anomaly) drops below a certain value, typically $1/e$ (Kuragano & Kamachi 2000, Hosoda & Kawamura 2004, De Benedetti & Moore 2017). It can be thought of as the typical scale of a coherent feature (such as a front or eddy), as captured by the data set. These length scales vary greatly between different environmental variables; they also vary based on the resolution of the observations. We hypothesized that DCLSs impact our ability to investigate habitat selection in marine organisms. To detect selection, animals would have to traverse length scales longer than the DCLS in a given region. We also postulate that the environmental selection of dynamic fields with very small DCLSs needs to be studied at a fine scale, and thus requires more precise estimates of animal location and higher-resolution environmental fields than the investigation of animals selecting for less dynamic environmental variables with larger DCLSs.

Here, we investigated how the interplay between animal location accuracy, selection strength, and DCLSs of environmental variables impacts our ability to statistically detect habitat selection by marine organisms. To do so, we created synthetic animal tracks with predetermined selection strengths for different environmental conditions. The specific environmental variables used were SST, chl *a* concentration, and a Lagrangian metric, finite-time Lyapunov exponent (FTLE), that captures the tendency of ocean currents

to aggregate or disperse passive tracers (Haller & Yuan 2000, Shadden et al. 2009). By comparing these synthetic tracks with generated pseudo-absences that mimic the movements of similar but environmentally naive individuals, we were able to determine what level of location accuracy is required to definitively identify selection for the studied environmental variables. In addition, we introduce the notion of ‘effective selection’ (combining both the test statistic and the significance level of Kolmogorov-Smirnov tests), significantly decreasing false-positive results in the absence of selection while only marginally increasing the rate of false-negative results at high sample sizes.

2. METHODS

2.1. Environmental variables and DCLS

Three different environmental fields were used in this study: SST, chl *a* concentration, and FTLE (Fig. 1A–C).

SST and chl *a* concentration were taken from MODIS-Aqua observations (JPL/OBPG/RSMAS 2020, NASA Goddard Space Flight Center et al. 2022). These fields have a native 9 km spatial resolution. The native time resolution is 1 d, but the data are processed so that each day consists of a backward rolling average

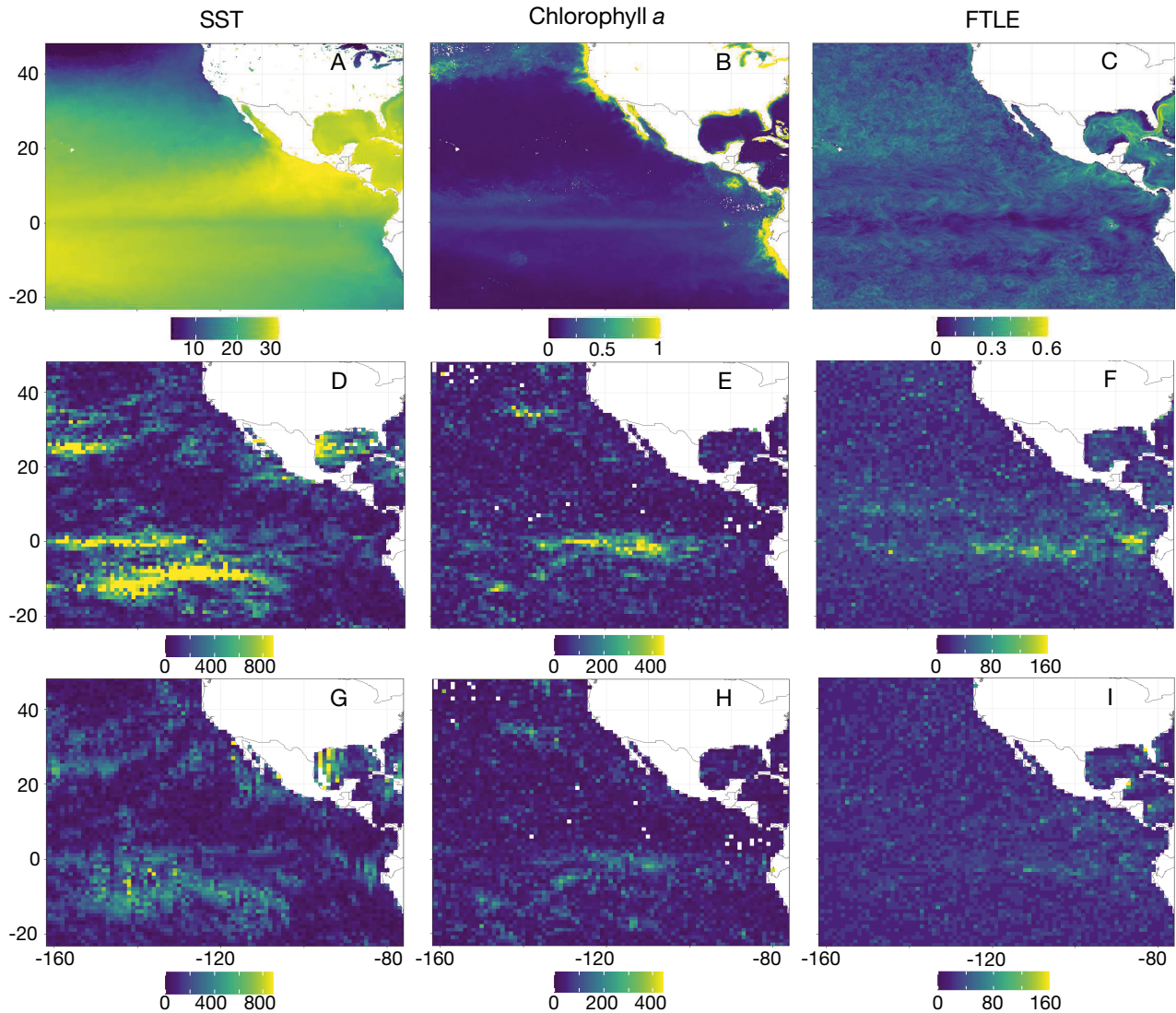


Fig. 1. Environmental conditions averaged over the period 1 May–19 July 2006, corresponding to the time period of the simulated tracks: (A) sea surface temperature (SST, in °C), (B) chlorophyll *a* (chl *a*) concentration (mg m⁻³), (C) finite-time Lyapunov exponent (FTLE, in d⁻¹). Zonal (east–west) decorrelation length scale (in km) for (D) SST, (E) chl *a* concentration, and (F) FTLE. Meridional (north–south) decorrelation length scale (in km) for (G) SST, (H) chl *a* concentration, and (I) FTLE. *x*- and *y*-axes of all panels are longitude and latitude, respectively

of the previous 8 d to increase data coverage, by decreasing missing data due to clouds and incomplete daily satellite coverage (Fig. S1).

The FTLE field is a tool for differentiating regions of the ocean subject to high dispersal from those prone to accumulation of passive tracers (e.g. Waugh et al. 2012, Peacock & Haller 2013, Sulman et al. 2013, Allshouse & Peacock 2015, Haller 2015, Hadjighasem et al. 2017, Callies & von Storch 2023). They measure relative dispersion, i.e. how far nearby water parcels separate or come together over a specified time interval. The FTLE field is a function of ocean currents, which were taken here from a global run of the Hybrid Coordinate Ocean Model (HYCOM), experiment 19.1, carried out by the Naval Research Laboratory at Stennis, MS, and archived by the HYCOM consortium at hycom.org. The FTLE field is computed using an along-trajectory velocity gradient integration on the model archive grid, which has a $1/12.5^\circ$ (~ 9 km) native resolution (Huntley et al. 2015).

We hypothesized that some marine organisms are attracted to regions of the ocean whose currents are conducive to accumulation of particles (e.g. plankton) over the recent past (Della Penna et al. 2015, Oliver et al. 2019, Lieber et al. 2023). Therefore, FTLEs were derived from an integration backward in time over 3 d. The sign convention adopted here has large positive FTLE values corresponding to highly attracting regions—water parcels that started far apart end up close together at the point in question and at the index time.

In addition, we also investigated the influence of data coverage by creating an environmental product corresponding to the FTLE product with the same data coverage as chl *a*. In practice, we removed FTLE data where chl *a* data were missing, effectively testing whether data gaps would affect our results.

For these 3 environmental variables (SST, chl *a*, and FTLE), we computed the zonal (in the east–west direction) and meridional (in the north–south direction) DCLS, defined as the *e*-folding scale of the variable anomalies (Hosoda & Kawamura 2004, De Benedetti & Moore 2017), i.e. the distance at which the correlation between 2 time series of the same variable drops below $1/e$. In practice, we started by subtracting the climatological signal (computed as the average for each calendar day over the period 2000–2010) from the daily signal. Then, for each reference point, we computed the correlation between the time series at that reference point and nearby time series. The distance at which the correlation drops below $1/e$ (in either the positive or negative direction) is taken as the DCLS at the reference point. The time series used

here were 80 d long, spanning from 1 May to 19 July 2006, which is the same time period as the one over which the animal tracks were generated (see Section 2.2).

2.2. Generation of synthetic animal tracks

For each of the 3 environmental variables, we created an array of 100 tracks (each 80 d long), hereafter referred to as synthetic tracks (Fig. S2). We varied the strength κ of environmental selection ($\kappa = 0$ for no selection, $\kappa = 0.25, 0.5, 0.75, 1, 2, 5, 10, 20$ for increasing selection strengths for high values of the target environmental variable) and the level of uncertainty σ assigned to the locations within these tracks. The standard deviations associated with the track locations are $\sigma = 0$ (exact location), 1, 10, 25, 50, and 111 km ($\sim 1^\circ$). Our total data set of synthetic tracks thus consisted of $9 \times 6 \times 3$ (9 different κ values and 6 different σ values for the 3 environmental variables used) sets of 100 tracks, i.e. 16 200 tracks.

The synthetic tracks of daily positions were generated as biased random walks, following Pinti et al. (2022), although here step lengths were not fixed, to better mimic the daily movements of marine organisms. For each track, a random starting location was picked in the Northeast Pacific, with latitude between 10 and 40° N and longitude between 140 and 130° W. Then, for each time step, the location of the highest value of the target variable within a 50 km radius was found, and the distance and bearing to that location were computed. The bearing of the actual step was then computed by modulating this bearing by a random angle drawn from a von Mises distribution with mean 0 and concentration κ (Fig. 2A). The step length is the distance to the location of highest value modulated by a random distance drawn from a normal distribution with mean 0 and standard deviation (Fig. 2B), except for $\kappa = 0$. The case $\kappa = 0$ mimics environmentally naive organisms, and to be sure that no information about the environment can be used for pseudo-absences generation, the step length was only drawn from a normal distribution with mean 50 km and standard deviation = 20 km.

2.3. Generation of pseudo-absences

Following Pinti et al. (2022), 3 different kinds of pseudo-absences were generated: Brownian motion, correlated random walks, and joint correlated random walks. Pinti et al. (2022) also explored using Lévy

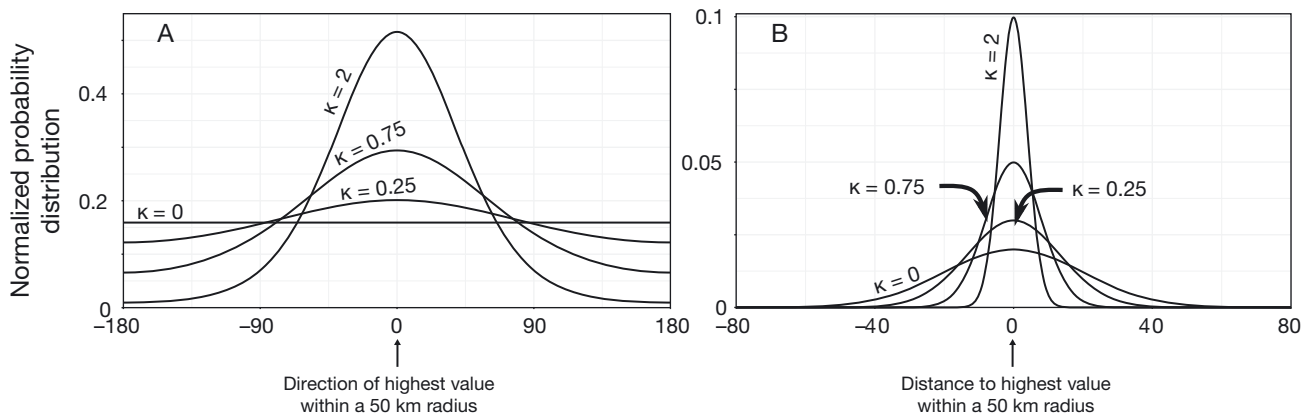


Fig. 2. (A) Von Mises distribution of relative bearing and (B) normal distribution of step length for movement models biased toward the highest value of the environmental variable within a 50 km radius for different values of the strength of environmental selection, κ

walks as a null model, but concluded that they could lead to high rates of false positive results — hence our use of only 3 different kinds of null models here. For each synthetic track, 100 tracks of each null model were generated, for a total of 300 pseudo-absences for each presence record from the synthetic tracks. Uncertainties associated with pseudo-absence locations are the same as the uncertainties associated with the generated synthetic locations.

Each pseudo-absence track starts at the starting location of its corresponding synthetic presence track, and is reset to the actual synthetic animal location at the beginning of every month, following the analysis of Pinti et al. (2022). Therefore, pseudo-absence tracks were maximum 31 d long. This ensured that pseudo-absences were constrained to the area around the real track and could still be reasonably considered to be in the same environment (Pinti et al. 2022).

Each null model generates locations based on step lengths and turning angles. The 3 null models differ only in the way they generate these step lengths and turning angles. For Brownian motion, the turning angle is drawn from a uniform distribution bound between -180 and 180° , and the step length is drawn from a normal distribution with mean and standard deviation equal to the mean and standard deviation of the step length distribution of the synthetic track. Correlated random walks and joint correlated random walks are simulated by drawing turning angles and step lengths directly from the empirical distribution of the synthetic track. The difference between these 2 methods is that for correlated random walks, step length and turning angles are drawn independently, whereas for joint correlated random walks, the

ordered pair (step length, turning angle) is drawn from a single step. Resulting pseudo-absence tracks for $\kappa = 1$ are pictured in Fig. S3 in the Supplement at www.int-res.com/articles/suppl/m732p001_supp.pdf.

2.4. Statistical analysis

Environmental data from the 3 variables were matched to each animal presence and pseudo-absence. When location uncertainty is >0 , however, it is not clear that the value at the recorded location is representative of the animal's environment. To account for this uncertainty in animal presence data, we averaged the environmental variables around each location, assuming a 2D Gaussian error distribution (i.e. the closer the observation is to the estimated location, the stronger the weight of this observation). The standard deviation for the distribution was set equal to the position uncertainty, hence accounting for the higher probability of having the animal in locations close to the most likely position estimate while still capturing the environmental variability within the range.

To test for evidence of selection, we performed 1-sided Kolmogorov-Smirnov (KS) tests, as implemented in 'ks.tests' of the R 'stats' package (R Core Team 2023). KS tests compare 2 cumulative distributions, with the test statistic D being the maximum distance between the 2 cumulative distributions. Here, the cumulative distributions correspond to the distribution of environmental variables matched to presence and pseudo-absence tracks, respectively. As such, D can be used as a proxy to estimate selection strength. The null hypothesis is that the cumulative

distribution of the target environmental variable for presences is 'not less than' (or 'not greater than') the cumulative distribution generated from pseudo-absences. For the 'not less than' test, if the null hypothesis is rejected, it means that the presence cumulative distribution function is below that of the pseudo-absence. The environmental variable distributions are shifted toward higher values, and animals select for high values of the environmental variable (e.g. higher temperatures or areas with higher chl *a* concentration) compared to environmentally naive organisms. Conversely, for the 'greater than' test, it means that animals select for lower values than environmentally naive organisms. Throughout this manuscript and unless specified otherwise, the significance level is set at $\alpha = 0.05$. Practically, very weak selection can, in many cases, not be distinguished statistically from no selection based on a finite sample (nor is it ecologically relevant). Therefore, we also imposed a threshold on the magnitude of the difference itself: we considered that animals are effectively selecting for higher (or lower) values of the environmental variable if the test result is significant ($p \leq 0.05$) and the test statistic D is > 0.05 . This threshold was chosen so as to decrease the rate of false-positive results in the absence of selection. This notion of effective selection allows us to discard false-positive results at high sample sizes in the absence of selection or at very weak selection strengths (see e.g. left columns of Fig. 5 and Fig. S7).

3. RESULTS

3.1. Tracks and environmental variables

The 3 environmental products considered (SST, chl *a* concentration, and FTLE) have different spatial characteristics (Fig. 1). First, the SST field is smoother than the chl *a* field, which is smoother than the FTLE field. In the Northeast Pacific, the DCLSs of the SST field vary between ~ 50 and 800 km, while the DCLSs of chl *a* and FTLE vary between 0 and 400 km and between 0 and 100 km, respectively. The median DCLS of the 3 products in the study region captures the differences in their spatial variability. SST has a median zonal DCLS of 130 km and a meridional DCLS of 111 km, while chl *a* has median zonal and meridional DCLSs of 46 and 47 km, respectively, and both FTLE DCLSs are 27 km. Generally, zonal DCLSs have higher maximum values than meridional DCLSs, even though the means are very similar for chl *a* and FTLE.

Because of the different spatial structures of these variables, the tracks generated based on the environmental selection of these fields are qualitatively different, especially at higher selection strengths (Fig. 3; for more samples, see Fig. S2). Tracks selecting for higher SST tend to be more spread and southward, while tracks strongly selecting for chl *a* and FTLE organize around ridges. Pseudo-absence tracks do not have a well-defined structure, as they are not cuing on any environmental variable (Fig. S3).

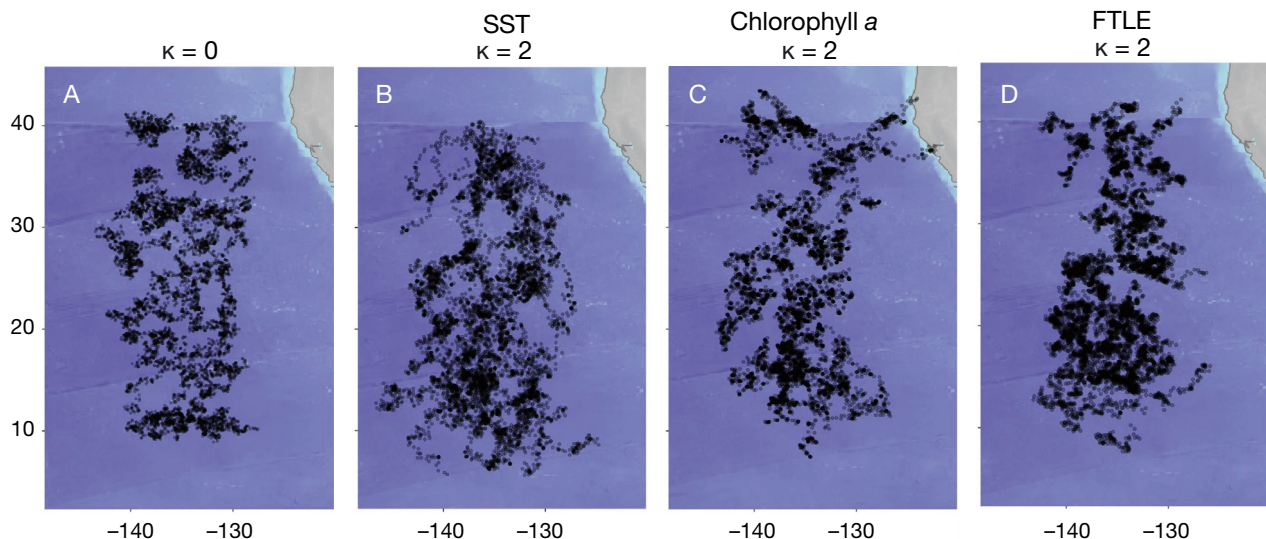


Fig. 3. Synthetic tracks of animals (A) selecting for no environmental variable ($\kappa = 0$), and strongly selecting ($\kappa = 2$) for (B) high sea surface temperature (SST), (C) high chlorophyll *a* concentration, and (D) high finite-time Lyapunov exponent (FTLE) values. There are 100 different 80 d tracks per panel. As the case $\kappa = 0$ corresponds to no environmental selection, the same set of tracks was used for all 3 environmental variables. x- and y-axes of all panels are longitude and latitude, respectively. Tracks for additional κ -values are shown in Fig. S2

The resulting distribution of SST, chl *a*, and FTLE for tracks strongly selecting for higher values of the environmental variables are shown in Fig. 4 (and histograms of environmental variable distributions for tracks with different selection strengths are plotted in Figs. S4–S6). For high selection strengths, as expected, the distribution of presence data is shifted towards higher values of the environmental variable compared to pseudo-absences. As κ decreases, the distribution of presence data shifts towards the distribution of pseudo-absences, all the way to $\kappa = 0$ where synthetic presence and pseudo-absence points have very similar distributions, consistent with the fact that this corresponds to animals not selecting for environmental variables. As σ increases (i.e. as location accuracy decreases), the overlap between distributions generated by animal presences and animal pseudo-absences increases, especially for FTLE and chl *a* concentration (Fig. 4). This is because the larger scale

averaging means that all values are more similar to each other, especially for environmental products with lower DCLS. The statistical analysis unravels the differences between these distributions as κ and σ vary.

3.2. Statistical analysis

To understand the interplay between position uncertainty σ , selection strength κ , and environmental variable DCLSs, we ran a total of 162 experiments (6 values of σ , 9 values of κ , 3 environmental fields with different DCLSs). In the following, we will focus on a selection of the results while describing the general patterns.

Fig. 5 shows the value of D for selection for higher values as a function of sample size (number of tracks). The corresponding data for selection for lower values is given in Fig. S7. As suggested by the probability density functions of environmental variables, the stronger the selection and the higher the location accuracy are, the higher is the test statistic. In addition, the FTLE field with gaps did not reveal results qualitatively different from the complete FTLE field (Figs. S7M–P & S8).

As the sample size increases, D decreases and converges to a point that depends on the environmental variable considered, the strength of the selection, and the location accuracy. In the absence of selection ($\kappa = 0$), some tests yield differences in distributions that are statistically significant results, yet with very small D (< 0.05).

A possibility to decrease these false-positive results would be to decrease the significance level below $\alpha = 0.05$. However, p-values usually decrease as sample sizes increase (Figs. S9–S16) and at large sample sizes, they can reach values as low as 10^{-5} in the absence of selection (e.g. Figs. S10O,P & S15M–P). Decreasing the level of significance to 10^{-5} would strongly decrease the rate of false-positive results in the absence of selection, but it would also consequently increase the rate of false-negative results in the presence of selection with limited sample size or with low accuracy. For example, setting the significance limit to 10^{-5}

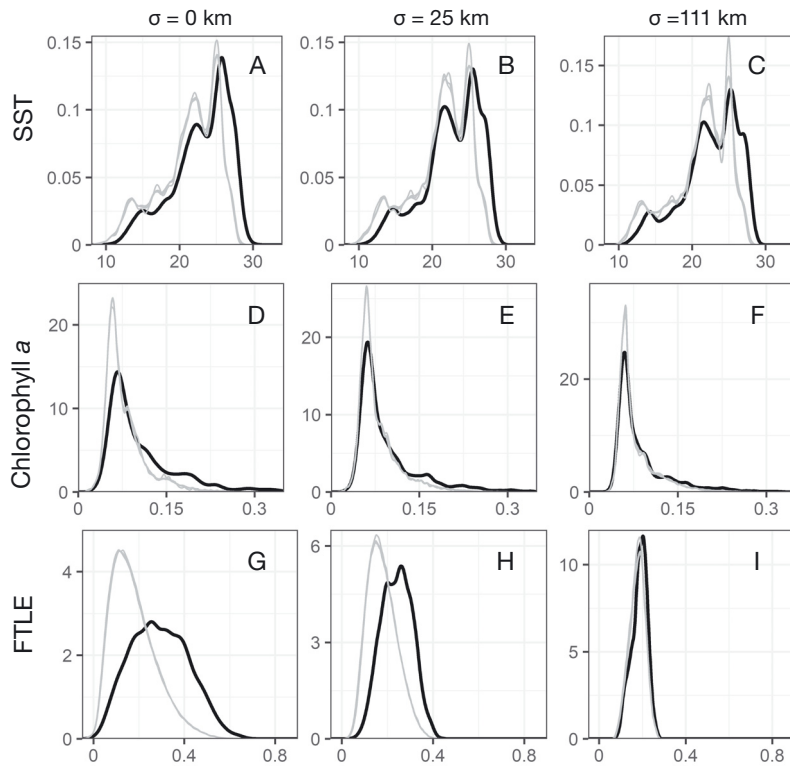


Fig. 4. Probability density functions of (A–C) sea surface temperature (SST, in $^{\circ}\text{C}$), (D–F) chlorophyll *a* concentration (in mg m^{-3}), and (G–I) finite-time Lyapunov exponent (FTLE, in d^{-1}) at synthetic presence (black) and pseudo-absence (grey) points, for presence tracks biased towards high values of the environmental variables ($\kappa = 2$). x-axes represent the environmental variables (SST, chlorophyll *a* concentration, or FTLE), y-axes represent the probability density functions. The first column (A,D,G) corresponds to locations known with perfect accuracy ($\sigma = 0$), the second column (B,E,H) to locations known with moderate accuracy ($\sigma = 25$ km), and the third column (C,F,I) to locations known with weak accuracy ($\sigma = 111$ km)

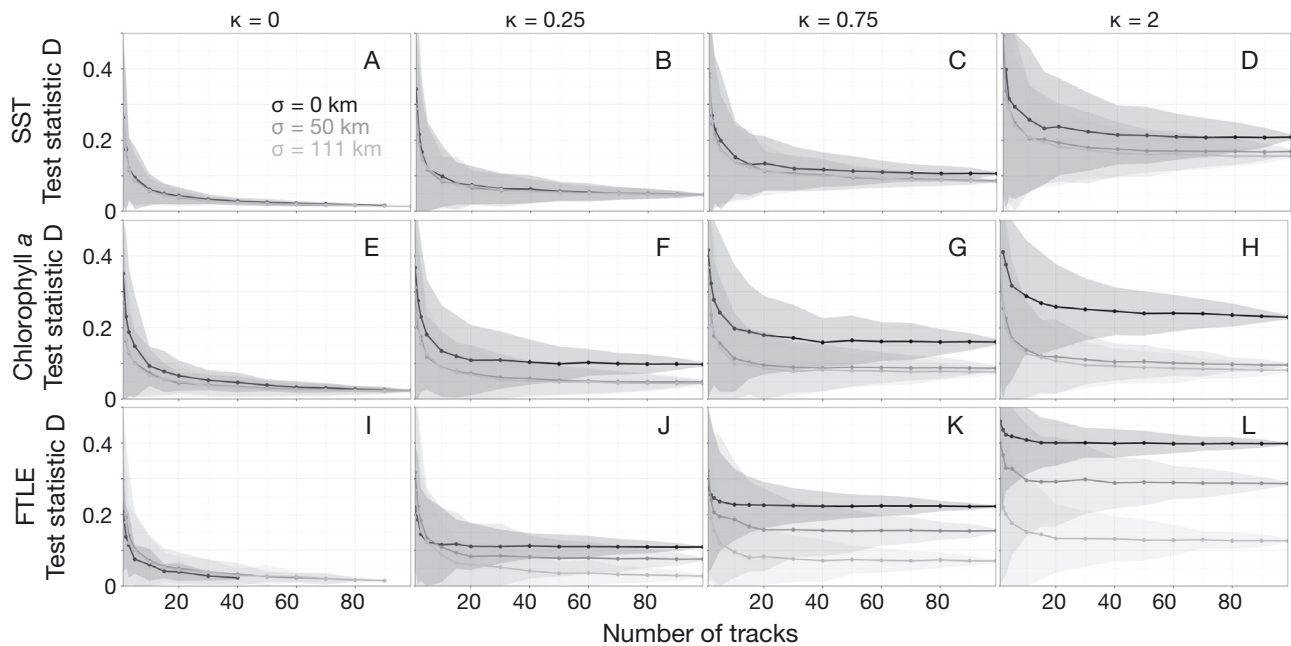


Fig. 5. Statistic D of tests of selection for higher values of (A–D) sea surface temperature (SST), (E–H) chlorophyll a , and (I–L) finite-time Lyapunov exponent (FTLE) at different selection strengths (κ , in columns) and for 3 different accuracies of geo-location estimates (σ , shades of grey). Shaded areas show the ensemble-mean estimate ± 3 standard deviation. Only test statistics of tests with $p < 0.05$ are included

would require more than 30 tracks to reliably detect selection for higher SST in the case of $\kappa = 0.25$, even when the tracks are reported with perfect accuracy (Fig. S9I). If the tracks have an uncertainty of 50 km, which is not unusual for animals tracked with ARGOS tags (Douglas et al. 2012), detecting selection for high chl a concentration requires more than 20 tracks in the case of strong selection ($\kappa = 2$ or more), and more than 80 tracks in the case of weak selection ($\kappa = 0.25$) (Fig. S10C,K). These very large sample sizes are very rare in ecological studies, and setting up such a stringent threshold would hinder our ability to detect selection in most practical cases. The case of $\kappa = 0$ with perfect position accuracy reveals that different animals following different movement models are likely to sample slightly different environments even without a selection bias. These very small differences show up even with very large sample sizes, but they are not ecologically relevant. Thus, we decided to evaluate effective selection here, i.e. selection with a p -value below 0.05, but with $D \geq 0.05$. This ensures that statistically significant results are not wrongly categorized as 'selecting' for higher (or lower) values of environmental variables, while still being able to detect selection when it is strong enough.

The ability to detect effective selection depends on sample size, selection strength, and location accuracy (Fig. 6; Figs. S17–S20 for selection for high values of

environmental variables, and Figs. S21–S25 for selection for low values of environmental variables). For FTLEs with and without data gaps (Fig. 6I–L; Fig. S26), the results are similar, suggesting that the method is not too sensitive to incomplete data coverage.

In the absence of selection, a large enough sample size allows us to confidently rule out effective selection (Fig. 6A,E,I). In the case of SST, using more than ~ 20 tracks (or 5 years of data) allows us to rule out effective selection irrespective of the location accuracy. For both chl a and FTLE, the sample size needed to rule out selection varies between 13 and 18 tracks (3–4 years of data) and more than 45 tracks (~ 10 years of data) depending on location accuracy (the more uncertain the track, the more data are needed). At the other extreme, when the selection is strong ($\kappa \geq 2$), it is possible to detect effective environmental selection even with high location uncertainty and limited sample size (Fig. 6D,H,L). As selection strength is weakened ($\kappa = 0.75$), large position uncertainty begins to have an impact on selection identification skill, at least for the variables with shorter DCLS (Fig. 6C,G,K). It is at weak selection strength ($\kappa = 0.25$) that the relationship between location accuracy, sample size, and identification skill becomes more complex (Fig. 6B,F,J). For chl a and FTLE, the general pattern of higher detection skills for larger sample sizes holds, except for cases with high position uncertainty.

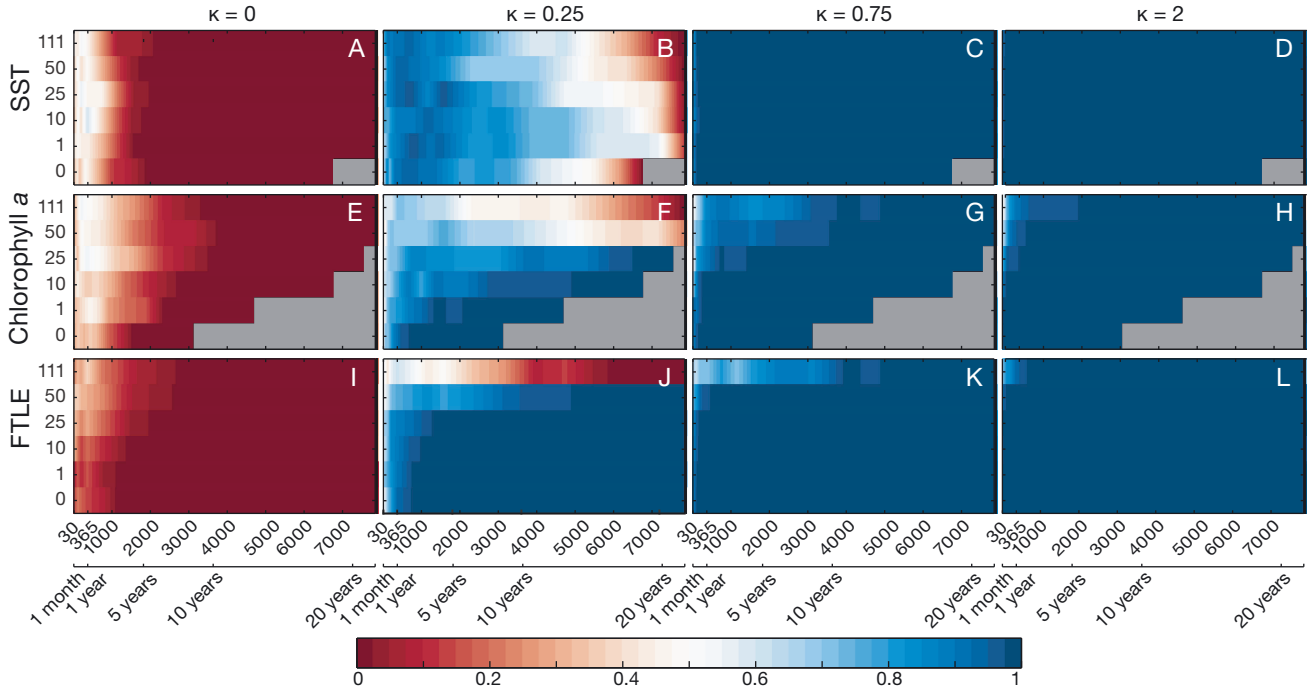


Fig. 6. Fraction of tests showing effective selection for higher values of (A–D) sea surface temperature (SST), (E–H) chlorophyll a , and (I–L) finite-time Lyapunov exponent (FTLE) at different selection strengths (κ , in columns), location accuracy (rows of each panel), and sample sizes (columns of each panel). Sample sizes are plotted as number of data points for which environmental data are available (in both number of data points and corresponding time span). At high location accuracy, we averaged the environmental variables over small areas, resulting in many track locations with missing data points for environmental products acquired with satellites. This lack of data is apparent in panels A–H, where grey patches correspond to missing data. For the case with perfect accuracy ($\sigma = 0$), we would need to generate approximately 20 000 days of tracking data to have 8000 days of tracking data with matching chl a concentration data

For SST, regardless of position uncertainty, Fig. 6B shows the counter-intuitive pattern that smaller sample sizes allow for a more reliable identification of effective selection bias. This is a direct result of the D thresholding (Fig. 5B,F,J): at small selection strength, the SST sample distributions for presence and pseudo-absence tracks differ only slightly for large sample sizes (and similarly for chl a and FTLE in cases with large position uncertainty). The differences are insufficient to be picked up as indicators of bias. The larger D -values for smaller sample sizes mean that our algorithm detects selection bias. However, we caution that these larger values are not an indication of the strength of the bias but an artefact of undersampling.

4. DISCUSSION

4.1. Detecting and quantifying environmental selection

In this paper, we explored our ability to detect marine animal environmental selection using KS tests, depending on a number of environmental and techni-

cal limitations. The ability to detect environmental selection depends on the DCLS of the environmental variable considered, on the selection strength of the animal, and on the sample size and accuracy of the tracking data set available. While we focused on SST, chl a , and FTLE in this study, our results are transferable to other environmental variables. Once the DCLSs of other variables are computed, one can refer to the variable with a similar DCLS in Fig. 6 to assess confidence in their selection results.

Setting a threshold on the test statistic D enables us to detect effective selection, thus considerably decreasing the risk of false-positive results while moderately decreasing our ability to detect very weak selection. We found that setting the D threshold to 0.05 and the significance threshold to 0.05 yields reliable selection detection (termed here effective selection), given enough data. The amount of data needed to have full confidence in a positive result depends on the DCLS of the investigated environmental variable, and can be defined as the amount of data necessary to fully rule out environmental selection in naive marine organisms (Fig. 6A,E,I). As an upper limit, more than 10 years of daily tracking data (~3650 daily location

estimates) was enough to have no false-positive results, no matter the DCLS of the environmental variables investigated here.

Another important result of this study is the possibility to quantify selection using the test statistic D as a proxy for selection strength. Indeed, for a given track accuracy, the stronger the selection strength is, the higher is the asymptotic D -value as sample size increases (Fig. 5). Provided with a large enough data set of animal tracks, performing KS tests on increasing fractions of the data set can reveal if the test statistic obtained for the entire data set is close to a converged value or not—and thus if the selection strength can be confidently quantified.

4.2. Different types of animal tracking data and their impact on environmental selection detection

A major discrepancy between real animal tracking data sets and the presence tracks generated here is that observed data can have varying accuracy and are not necessarily evenly spaced in time. Processing the data with a state–space model allows us to resample the track evenly (Jonsen et al. 2005). However, state–space models usually assume that animals follow correlated random walks without accounting for environmental selection, thereby biasing processed data against selection detection. In addition, the spatial uncertainties associated with processed points temporally far away from observed points is usually greater, further increasing the range of location uncertainties within tracks. As we assumed here that location uncertainty was homogeneous within the entire data set, adapting this method to real animal locations may require discarding location estimates that are too uncertain.

This consideration is particularly important for more uncertain tracking methods. Fastloc GPS tags provide accuracy within 700 m and most of the time within 50 m (Dujon et al. 2014), well below the resolution of the environmental products considered here. However, ARGOS and PSATs have larger error ranges. ARGOS tags provide locations with a quality location class (Douglas et al. 2012), allowing the filtering of data to keep only the most precise—at the expense of sample size (Thomson et al. 2017). PSATs, because they mostly geolocate animals thanks to light levels and SST (even though other variables such as bathymetry, magnetic field, or temperature profiles can be used; Braun et al. 2018, Nielsen et al. 2019, 2020), have an even wider range, with errors in actual positions routinely around 1° in longitude and

latitude (Wilson et al. 2007). Yet, for highly migratory marine species, it appears that PSATs can perform better than ARGOS tags when building species distribution models (using methods other than ours, such as generalized linear or additive mixed models, or boosted regression trees) using variables with high DCLS (Braun et al. 2023). This highlights the importance of considering the scale of the question being investigated. Following the results of this study, we recommend using either ARGOS or GPS tag records when investigating environmental selection in marine organisms for variables with low DCLS. In general, generating the most accurate location data possible is often the safest way to get reliable results. In particular, when limited to PSAT data because of the ecology of the animal, relying on manufacturer-provided geolocation models often yields inaccurate results relative to more advanced geolocation models (Braun et al. 2018, Nielsen et al. 2023). More precise locations will naturally lead to more accurate ecological results, and will also enable investigating ecological questions requiring more precise data, such as the selection of environmental variables with comparatively lower DCLS than would be possible without these data.

4.3. Scale, resolution, and accuracy of environmental variables

In addition to the resolution at which animal locations are observed, the scale at which we acquire oceanographic data is important when investigating environmental selection. The data sets we used here have a fixed 9 km resolution (8 km in the case of FTLE), smoothing variability at smaller scales and making sub-mesoscale features undetectable. Spatial resolution can be finer if environmental data are acquired differently. For example, Lagrangian coherent structures have been computed from high-frequency radars from 6 km down to 500 m depending on the frequency used (Kim et al. 2011, Berta et al. 2014, Oliver et al. 2019), but usually at the expense of spatial coverage. Spatially averaging the environmental data increases spatial coverage, creating a trade-off between data coverage and spatial resolution. In addition to technical considerations regarding this trade-off, this raises ecological questions as to what scale is appropriate when investigating selection of marine organisms. A larger scale may be necessary to have a good spatial coverage, but may blur small variations that animals may use and select for. As it is not possible to detect selection happening at a

finer scale than the resolution of the data, it is important to match the scale at which animals can move and the scale at which we can acquire environmental data as well as possible.

The way oceanographic data are acquired dictates their resolution, but also their accuracy. While data observed *in situ* (and, to a lesser extent, remotely sensed data) have a relatively strong spatio-temporal accuracy, data generated by models may have lower accuracy. For example, the FTLEs used here were computed from HYCOM model outputs and not directly observed. Several studies have found that the uncertainties in predictions of trajectories and Lagrangian coherent structures computed from HYCOM model outputs are, on average, 50 km, with outliers as large as 100 km (Huntley et al. 2011, Muscarella et al. 2015, Thoppil et al. 2021). Here, this discrepancy is irrelevant as the tracks are synthetic and generated using the model FTLE outputs directly, but it is of importance when dealing with real tracks of animals that are cuing on actual Lagrangian structures. Incorporating the environmental data uncertainty in this method will likely have the same effect as considering the track uncertainty — it will average out specific values that animals may cue on with background conditions, thus decreasing our ability to detect selection.

Data coverage is another factor that might influence our ability to detect selection. The amount of missing data in this study differed for the 3 environmental fields (Fig. S1). As the FTLE field is a model output, there were no data gaps, while there were some gaps in the SST and even more in the chl a fields. Thus, not all points could be matched to all environmental variables (Table 1). While the same number of tracks were used with all 3 variables, this translates into a different number of data points with matched environmental variables, with FTLE having the most data points and chl a the fewest. The number of data points also increases as the assumed accuracy

Table 1. Fraction (%) of animal presence location estimates with environmental data for different location standard error (σ). Each cell of this table corresponds to 72 000 data points (80 d tracks [$n = 100$]) for 9 different values of the strength of environmental selection, (κ). SST: sea surface temperature; FTLE: finite-time Lyapunov exponent

	σ					
	0 km	1 km	10 km	25 km	50 km	111 km
Chl a	40.7	67.3	87.7	96.2	99.6	100
SST	86.5	95.5	99.6	99.9	100	100
FTLE	100	100	100	100	100	100

of location estimates decreases, as less precise estimates mean that the average is performed across a larger area and thus more likely to encompass existing environmental data.

4.4. Increasing sample size: beware of heterogeneity

Larger animal tracking data sets lead to improved confidence in selection detection. Therefore, it is tempting to aggregate data from different sources, geographic locations, time of the year, or even from different years to increase sample size. However, disregarding the variability that can exist in data sets may yield to inaccurate results. Populations from different geographic areas may experience a different range of environmental conditions. Thus, different populations might be adapted to and select for different environmental conditions — such as shark species living in pelagic environments and very clear waters in the Pacific Ocean but near the coasts in turbid environments in the Atlantic Ocean (Merson & Pratt 2001, Papastamatiou et al. 2006). Animals may change behavior and selection strategy as they grow (ontogenetic niche partitioning, Grubbs 2010) or throughout the year, for example when they migrate to colder, lower latitudes (Horton et al. 2011), or when they move between offshore and inshore grounds with different productivity levels (Weng et al. 2007). Finally, conditions may change over time, for example as a result of climate change, resulting in changing experienced environmental conditions for organisms, such as a decrease in available prey (Meyer-Gutbrod et al. 2023).

Depending on the flexibility of animal movements, these longer-term, multi-year changes in environmental conditions may be chosen or imposed on marine organisms. Migrating organisms can follow specific cues and select for particular environmental conditions, or they can use memory to reproduce movement patterns of previous years, potentially resulting in a mismatch with their optimal environmental conditions (Abrahms et al. 2019, Fagan 2019). How the changing baseline will impact environmental selection in marine organisms is yet to be determined and will depend on how much animals rely on memory vs. environmental cues. To this end, it would be useful to compare animals tracks not only to instantaneous environmental products but also to past variables and variables averaged at different temporal scales to understand the importance of conditions experienced in the past in shaping marine animals' movements and environmental niches.

5. CONCLUSION

The accuracy of animal location and the spatial variability in environmental variables (measured through DCLSs) impact our ability to detect selection in marine organisms. As previously shown (Pinti et al. 2022), increasing sample size allows for more robust detection of environmental selection, except when selection is very weak. Setting a significance level at 0.05 and a cumulative distribution difference threshold level at 0.05 to detect effective selection in marine organisms significantly reduces the risk of false-positive results, while only increasing the risk of false-negative results in the case of very weak selection strength.

The amount and accuracy of tracking data needed to detect selection depends on the DCLS of the environmental variable tested. In practice, data acquired with Fastloc GPS (and ARGOS) tags are the most precise and thus the most efficient to use when it comes to detecting and quantifying selection strength, even though a large fraction of marine organisms do not surface and therefore cannot be studied with such technology.

Finally, it is important to mention that we are detecting correlation but not necessarily causation. While we can confidently say whether organisms preferentially associate with certain environmental variables thanks to this method, we cannot ascertain if they actively target these variables or variables correlated to them, or whether they target these conditions because of immediate cues or because of memory and movement patterns acquired through social learning. Testing for environmental selection with past environmental conditions and at different temporal scales might help answer this question.

Data availability. All code used to perform the analysis is available at https://github.com/JeromeAqua/Selection_accuracy/.

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LITERATURE CITED

- Abrahms B, Hazen EL, Aikens EO, Savoca MS and others (2019) Memory and resource tracking drive blue whale migrations. *Proc Natl Acad Sci USA* 116:5582–5587
- Allshouse MR, Peacock T (2015) Refining finite-time Lyapunov exponent ridges and the challenges of classifying them. *Chaos* 25:087410
- Berta M, Bellomo L, Magaldi MG, Griffa A and others (2014) Estimating Lagrangian transport blending drifters with HF radar data and models: results from the TOSCA experiment in the Ligurian Current (North Western Mediterranean Sea). *Prog Oceanogr* 128:15–29
- Block BA, Costa DP, Boehlert GW, Kochevar RE (2002) Revealing pelagic habitat use: the tagging of Pacific pelagics program. *Oceanol Acta* 25:255–266
- Braun CD, Galuardi B, Thorrold SR (2018) HMMoce: an R package for improved geolocation of archival-tagged fishes using a hidden Markov method. *Methods Ecol Evol* 9:1212–1220
- Braun CD, Arostegui MC, Farchadi N, Alexander M and others (2023) Building use-inspired species distribution models: using multiple data types to examine and improve model performance. *Ecol Appl* 33:e2893
- Callies U, von Storch H (2023) Extreme separations of bottle posts in the southern Baltic Sea — tentative interpretation of an experiment-of-opportunity. *Oceanologia* 65: 410–422
- De Benedetti M, Moore GW (2017) Impact of resolution on the representation of precipitation variability associated with the ITCZ. *Geophys Res Lett* 44:12519–12526
- Della Penna A, De Monte S, Kestenare E, Guinet C, D’Ovidio F (2015) Quasi-planktonic behavior of foraging top marine predators. *Sci Rep* 5:18063
- Douglas DC, Weinzierl RC, Davidson S, Kays R, Wikelski M, Bohrer G (2012) Moderating Argos location errors in animal tracking data. *Methods Ecol Evol* 3:999–1007
- Dujon AM, Lindstrom RT, Hays GC (2014) The accuracy of Fastloc-GPS locations and implications for animal tracking. *Methods Ecol Evol* 5:1162–1169
- Dulvy NK, Sadovy Y, Reynolds JD (2003) Extinction vulnerability in marine populations. *Fish Fish* 4:25–64
- Fagan WF (2019) Migrating whales depend on memory to exploit reliable resources. *Proc Natl Acad Sci USA* 116: 5217–5219
- Fahlbusch JA, Czapanskiy MF, Calambokidis J, Cade DE, Abrahms B, Hazen EL, Goldbogen JA (2022) Blue whales increase feeding rates at fine-scale ocean features. *Proc R Soc B* 289:20221180
- Grubbs RD (2010) Ontogenetic shifts in movements and habitat use. In: Carrier JF, Musick JA, Heithaus MR (eds) *Sharks and their relatives II: biodiversity, adaptive physiology, and conservation*. CRC Press, Boca Raton, FL, p 319–350
- Gunn J, Polacheck T, Davis T, Sherlock M, Betlehem A (1994) The development and use of archival tags for studying the migration, behavior and physiology of southern bluefin tuna, with an assessment of the potential for transfer of technology to groundfish research. *Proc ICES Symp Fish Migration* 21:1–23
- Hadjighasem A, Farazmand M, Blazeovski D, Froyland G, Haller G (2017) A critical comparison of Lagrangian methods for coherent structure detection. *Chaos* 27:053104
- Haller G (2015) Lagrangian coherent structures. *Annu Rev Fluid Mech* 47:137–162
- Haller G, Yuan G (2000) Lagrangian coherent structures and mixing in two-dimensional turbulence. *Physica D* 147: 352–370
- Halpern BS, Walbridge S, Selkoe KA, Kappel CV and others (2008) A global map of human impact on marine ecosystems. *Science* 319:948–952
- Hazen EL, Abrahms B, Brodie S, Carroll G, Welch H, Bograd SJ (2021) Where did they not go? Considerations for generating pseudo-absences for telemetry-based habitat models. *Mov Ecol* 9:5
- Horton TW, Holdaway RN, Zerbini AN, Hauser N, Garrigue

- C, Andriolo A, Clapham PJ (2011) Straight as an arrow: Humpback whales swim constant course tracks during long-distance migration. *Biol Lett* 7:674–679
- ✦ Hosoda K, Kawamura H (2004) Global space-time statistics of sea surface temperature estimated from AMSR-E data. *Geophys Res Lett* 31:L17202
- ✦ Huntley HS, Lipphardt BL, Kirwan AD (2011) Lagrangian predictability assessed in the East China Sea. *Ocean Model* 36:163–178
- ✦ Huntley HS, Lipphardt BL, Jacobs G, Kirwan AD (2015) Clusters, deformation, and dilation: diagnostics for material accumulation regions. *J Geophys Res Oceans* 120:6622–6636
- ✦ Hussey NE, Kessel ST, Aarestrup K, Cooke SJ and others (2015) Aquatic animal telemetry: a panoramic window into the underwater world. *Science* 348:1255642
- ✦ Jonsen I, Flemming JM, Myers R (2005) Robust state-space modeling of animal movement data. *Ecology* 86:2874–2880
- ✦ JPL/OBPG/RSMAS (2020) GHRSSST Level 2P global sea surface skin temperature from the Moderate Resolution Imaging Spectroradiometer (MODIS) on the NASA Aqua satellite (GDS2). MODIS Aqua L2P swath SST data set ver 2019.0. https://podaac.jpl.nasa.gov/dataset/MODIS_A-JPL-L2P-v2019.0
- Kim SY, Terrill EJ, Cornuelle BD, Jones B and others (2011) Mapping the U.S. West Coast surface circulation: a multi-year analysis of high-frequency radar observations. *J Geophys Res Oceans* 116:C03011
- ✦ Kuragano T, Kamachi M (2000) Global statistical space-time scales of oceanic variability estimated from the TOPEX/POSEIDON altimeter data. *J Geophys Res Oceans* 105:955–974
- ✦ Lee KA, Butcher PA, Harcourt RG, Patterson TA and others (2021) Oceanographic conditions associated with white shark (*Carcharodon carcharias*) habitat use along eastern Australia. *Mar Ecol Prog Ser* 659:143–159
- ✦ Lieber L, Füchtencordsjürgen C, Hilder RL, Revering PJ, Siekmann I, Langrock R, Nimmo-Smith WAM (2023) Selective foraging behavior of seabirds in small-scale slicks. *Limnol Oceanogr Lett* 8:286–294
- ✦ Lotze HK, Lenihan HS, Bourque BJ, Bradbury RH and others (2006) Depletion, degradation, and recovery potential of estuaries and coastal seas. *Science* 312:1806–1809
- ✦ Matley JK, Klinard NV, Barbosa Martins AP, Aarestrup K and others (2022) Global trends in aquatic animal tracking with acoustic telemetry. *Trends Ecol Evol* 37:79–94
- ✦ Merson RR, Pratt HL Jr (2001) Distribution, movements and growth of young sandbar sharks, *Carcharhinus plumbeus*, in the nursery grounds of Delaware Bay. *Environ Biol Fishes* 61:13–24
- ✦ Meyer-Gutbrod EL, Davies KTA, Johnson CL, Plourde S and others (2023) Redefining North Atlantic right whale habitat-use patterns under climate change. *Limnol Oceanogr* 68:S71–S86
- ✦ Møller AP, Rubolini D, Lehikoinen E (2008) Populations of migratory bird species that did not show a phenological response to climate change are declining. *Proc Natl Acad Sci USA* 105:16195–16200
- ✦ Muscarella P, Carrier MJ, Ngodock H, Smith S, Lipphardt BL Jr, Kirwan AD Jr, Huntley HS (2015) Do assimilated drifter velocities improve Lagrangian predictability in an operational ocean model? *Mon Weather Rev* 143:1822–1832
- ✦ NASA Goddard Space Flight Center, Ocean Ecology Laboratory, Ocean Biology Processing Group (2022) Moderate-resolution Imaging Spectroradiometer (MODIS) Aqua Ocean Color Data. <https://oceancolor.gsfc.nasa.gov/about/missions/aqua/>
- ✦ Nielsen JK, Mueter FJ, Adkison MD, Loher T, McDermott SF, Seitz AC (2019) Effect of study area bathymetric heterogeneity on parameterization and performance of a depth-based geolocation model for demersal fishes. *Ecol Model* 402:18–34
- ✦ Nielsen JK, Mueter FJ, Adkison MD, Loher T, McDermott SF, Seitz AC (2020) Potential utility of geomagnetic data for geolocation of demersal fishes in the North Pacific Ocean. *Anim Biotelem* 8:17
- ✦ Nielsen JK, Bryan DR, Rand KM, Arostegui MC, Braun CD, Galuardi B, McDermott SF (2023) Geolocation of a demersal fish (Pacific cod) in a high-latitude island chain (Aleutian Islands, Alaska). *Anim Biotelem* 11:29
- ✦ Oliver MJ, Kohut JT, Bernard K, Fraser W and others (2019) Central place foragers select ocean surface convergent features despite differing foraging strategies. *Sci Rep* 9:157
- ✦ Papastamatiou YP, Wetherbee BM, Lowe CG, Crow GL (2006) Distribution and diet of four species of carcharhinid shark in the Hawaiian Islands: evidence for resource partitioning and competitive exclusion. *Mar Ecol Prog Ser* 320:239–251
- ✦ Peacock T, Haller G (2013) Lagrangian coherent structures: the hidden skeleton of fluid flows. *Phys Today* 66:41–47
- Peterson C, Lubchenco J (1997) Marine ecosystem services. In: Daily GC (ed) *Nature's services: societal dependence on natural ecosystems*. Island Press, Washington DC, p 177–194
- ✦ Pinti J, Shatley M, Carlisle A, Block BA, Oliver MJ (2022) Using pseudo-absence models to test for environmental selection in marine movement ecology: the importance of sample size and selection strength. *Mov Ecol* 10:60
- ✦ Pörtner HO, Knust R (2007) Climate change affects marine fishes through the oxygen limitation of thermal tolerance. *Science* 315:95–97
- R Core Team (2023) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna
- ✦ Scales KL, Hazen EL, Jacox MG, Edwards CA, Boustany AM, Oliver MJ, Bograd SJ (2017) Scale of inference: on the sensitivity of habitat models for wide-ranging marine predators to the resolution of environmental data. *Ecography* 40:210–220
- ✦ Shadden SC, Lekien F, Paduan JD, Chavez FP, Marsden JE (2009) The correlation between surface drifters and coherent structures based on high-frequency radar data in Monterey Bay. *Deep Sea Res II* 56:161–172
- ✦ Sibert JR, Musyl MK, Brill RW (2003) Horizontal movements of bigeye tuna (*Thunnus obesus*) near Hawaii determined by Kalman filter analysis of archival tagging data. *Fish Oceanogr* 12:141–151
- ✦ Sulman MH, Huntley HS, Lipphardt BL, Jacobs G, Hogan P, Kirwan AD (2013) Hyperbolicity in temperature and flow fields during the formation of a Loop Current ring. *Non-linear Process Geophys* 20:883–892
- ✦ Tew Kai E, Rossi V, Sudre J, Weimerskirch H and others (2009) Top marine predators track Lagrangian coherent structures. *Proc Natl Acad Sci USA* 106:8245–8250
- ✦ Thomson JA, Börger L, Christianen MJ, Esteban N, Laloë JO, Hays GC (2017) Implications of location accuracy and data volume for home range estimation and fine-

- scale movement analysis: comparing Argos and Fastloc-GPS tracking data. *Mar Biol* 164:204
- ✦ Thoppil PG, Frolov S, Rowley CD, Reynolds CA and others (2021) Ensemble forecasting greatly expands the prediction horizon for ocean mesoscale variability. *Commun Earth Environ* 2:89
- ✦ Waugh DW, Keating SR, Chen ML (2012) Diagnosing ocean stirring: comparison of relative dispersion and finite-time Lyapunov exponents. *J Phys Oceanogr* 42:1173–1185
- ✦ Weng KC, Boustany AM, Pyle P, Anderson SD, Brown A, Block BA (2007) Migration and habitat of white sharks (*Carcharodon carcharias*) in the eastern Pacific Ocean. *Mar Biol* 152:877–894
- ✦ Wilson SG, Stewart BS, Polovina JJ, Meekan MG, Stevens JD, Galuardi B (2007) Accuracy and precision of archival tag data: a multiple-tagging study conducted on a whale shark (*Rhincodon typus*) in the Indian Ocean. *Fish Oceanogr* 16:547–554

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