



Understanding organism—habitat relationships and critically evaluating reference areas is key to marine protected area assessment

Conner Jainese*, Peter M. Carlson, Katelin Seeto, Lyndsey McNeill, Kelly Sivertson, Jennifer E. Caselle

Marine Science Institute, University of California Santa Barbara, Santa Barbara, CA 93106-6150, USA

ABSTRACT: Marine protected areas (MPAs) are specified ocean areas designated by management agencies in which extractive human activities (i.e. fishing) are either prohibited ('no-take') or limited in some way. MPAs have been implemented globally as a conservation tool to improve the health and function of marine ecosystems. Research has focused on assessing MPA effectiveness; however, certain habitats and communities are often avoided because they are difficult or expensive to monitor. Mesophotic (30–100 m) rocky reef fish communities are valuable commercial and recreational resources that are highly targeted but often overlooked in monitoring due to depthrestricted sampling. We used 2 MPAs in the statewide protection network of California (USA) that prohibit fishing for these species, along with paired reference sites, to test how protection status along with environmental conditions influenced the abundance and biomass of 3 highly targeted species groups with varying life histories and habitat preferences. Depth and habitat were strong predictors for all groups: ocean whitefish Caulolatilus princeps, California sheephead Semicossyphus pulcher, and targeted rockfish (Sebastes spp.). However, the pattern of these effects differed between the species groups, and the influence of protection was mixed. This work highlights how species with high habitat affinities benefit differently from protection, as a function of depth and habitat representation within the MPA/reference pair. To accurately evaluate MPAs, and the network as a whole, researchers must recognize organism-habitat relationships and incorporate them when assessing conservation efforts.

KEY WORDS: Baited remote underwater video \cdot Mesophotic \cdot Marine protected area \cdot Habitat preference \cdot Marine conservation \cdot California \cdot Fish ecology

1. INTRODUCTION

Marine protected areas (MPAs) have been implemented globally as a means of protecting marine ecosystems from anthropogenic pressures (Gleason et al. 2013, Maestro et al. 2019, Sullivan-Stack et al. 2022). The primary goal of most MPAs is to directly alleviate fishing pressure and other human impacts, with the expected outcome of conserving and maintaining marine biodiversity. Other benefits, such as increased catch rates outside of protection zones and economic stability for local communities, are also predicted to occur (Murray et al. 1999, Sala & Giakoumi 2018). There is strong evidence that an MPA network, as opposed to a single MPA, is more effective at broadly protecting a diverse suite of species across various life stages and habitats (Gaines et al. 2010, Grorud-Colvert et al. 2014). Well-designed MPA networks are able to incorporate nursery areas, account for ontogenetic habitat shifts, recognize different species habitat associations, and protect vulnerable spawning grounds (Grüss et al. 2014, Olson et al. 2019). Monitoring all of these habitats is difficult, and it is rare that all habitats within a single MPA or across an MPA net-

^{*}Corresponding author: connerjainese@ucsb.edu

[©] The authors 2024. Open Access under Creative Commons by Attribution Licence. Use, distribution and reproduction are unrestricted. Authors and original publication must be credited.

work are effectively monitored. Monitoring priority is often given to those habitats more easily or cheaply accessed (shallow-water reefs, grass beds, shorelines), while more challenging habitats are often datapoor (Field et al. 2007, Noble-James et al. 2023).

Mesophotic (30-100 m) reefs represent one habitat that has been historically difficult to monitor, and a growing body of research acknowledges our relatively incomplete understanding of these communities globally (James et al. 2017, Kahng et al. 2017, Cerrano et al. 2019). Mesophotic reefs support a diverse suite of commercially valuable species, and it is estimated that a significant portion of commercial groundfish harvest falls within this depth zone (Miller et al. 2017, Jacquemont et al. 2024). As a result, these fishes are under-surveyed and relatively data-poor, while simultaneously being highly targeted. Along the west coast of California (USA), many of the mesophotic fisheries are actively managed, with enforcement agencies frequently revising depth restrictions and species-specific take limits. Active management requires monitoring data from these depths, and a combination of fisheries-dependent and independent data has been shown to provide a robust measure of stock size (Dennis et al. 2015, Howard et al. 2023). Baited remote underwater video (BRUV) systems have opened new opportunities for better understanding of mesophotic reefs in temperate and tropical ecosystems by providing a fisheries-independent, non-destructive sampling technique that can operate at depths beyond recreational or even most scientific SCUBA diving limits (Brokovich et al. 2008, Andradi-Brown et al. 2016, Brown et al. 2022).

Mesophotic reefs represent an ecologically important habitat that contains distinct invertebrate and fish communities (Williams et al. 2019, Honeyman et al. 2023, Bell et al. 2024). Below 30 m depth, solar irradiance begins to attenuate, limiting photosynthesis and algal growth that characterizes shallower reefs. At these depths, structure-forming sessile invertebrates, many themselves of conservation concern, such as gorgonians, anemones, calcareous algae, and bryozoans, dominate the benthos (Ponti et al. 2018). In the northern hemisphere, the mesophotic temperate reef fish community is dominated by rockfishes (genus Sebastes), a diverse assemblage of ecologically and economically important fishes that includes >100 recognized species (Love et al. 2002, Love & Yoklavich 2006, Love 2011). The mesophotic depth zone offers a relatively stable set of environmental conditions that can function as nursery habitat and a refuge space for species that were historically found along wider depth gradients (Baillon et al. 2012, Kahng et al. 2017,

Giraldo-Ospina et al. 2020). These habitats also provide an important ontogenetic bridge between shallow nursery habitats and deeper adult habitats (Love 2011, Swadling et al. 2022). Finally, these deep habitats might be critical refuges from the effects of climate change, such as increases in sea water temperature (MacDonald et al. 2016, Pereira et al. 2018). Together these factors make mesophotic reefs important contributors to marine biodiversity and a necessary component of an effective MPA network.

One of the expected benefits of an MPA is the buildup of targeted species within its boundaries and consequently, the performance of an MPA is often measured by its ability to enhance the biomass and abundance of fisheries targets (Caselle et al. 2015, Lenihan et al. 2022). Recreational and commercial fisheries often have a number of targeted species that utilize a wide breadth of habitats across the seascape (California Department of Fish and Game Marine Region 2003, Scholz et al. 2006). Effective MPA networks should protect habitats ranging from sand, sand-rock ecotone, rocky reefs, and pelagic zones, while incorporating these habitats across an extensive depth range (Young & Carr 2015). Despite the fact that MPAs are an ecosystem conservation tool, it is unlikely that any MPA can achieve a 'one size fits all' design; instead, any single MPA will likely be more effective at protecting some species compared to others. Factors such as species movement, the availability of preferred habitat, oceanographic conditions, predator and prey abundances, and recruitment dynamics will all influence the ability of an MPA to enhance particular targeted species (Moffitt et al. 2011).

With the purpose of better understanding the effects of MPAs on mesophotic rocky reef fish communities, we implemented the use of BRUV camera systems at 2 geographically distinct MPAs (and nearby reference areas) within California's Northern Channel Islands (Fig. 1): Carrington Point State Marine Reserve (SMR) at Santa Rosa Island and Anacapa Island SMR/State Marine Conservation Area (SMCA). Straddled across a biogeographic transition zone, where cool southernflowing waters of the California Current meet warmer northern-flowing waters from the Southern California Counter Current (Horn & Allen 1978), these 2 MPAs are characteristically distinct, occupying opposite ends of a strong environmental gradient that exists across the Northern Channel Islands. The primary variation along this gradient is in sea surface temperature (SST), accompanied by other related factors including productivity, frequency of disturbance/ wave exposure, algal species persistence, and level of human interaction (Harms & Winant 1998, Hamilton



Fig. 1. Study region and the 2 focal marine protected areas (MPAs) in the Santa Barbara Channel, Northern Channel Islands (NCI), California, USA. (A) Location of the NCI along the West Coast of North America. (B) NCI, with red polygons delineating the NCI MPAs within the state-wide California Marine Protected Area Network. (C) Carrington Point MPA and associated reference area. (D) Anacapa MPA complex and associated reference area. In panels (C) and (D), dark grey shading represents mapped hard bottom, and 10 m bathymetry lines are light grey. Locations of baited remote underwater video (BRUV) surveys by year are shown in Fig. S2 in the Supplement at www.int-res.com/articles/suppl/m751p115_supp.pdf

et al. 2010, Caselle et al. 2015). Additionally, these 2 MPAs were selected because they contain appropriate rocky habitat within our study depth zone; they provide complete protection ('no-take') for the demersal fish communities on which this study focuses; and both MPAs have a number of other ongoing, complementary MPA monitoring projects (Honeyman et al. 2023).

We utilized stereo BRUV surveys because they are well suited for capturing fish community data across this under-sampled depth range and are useful for a variety of habitat types (Cappo et al. 2004, Heyns-Veale et al. 2016, Whitmarsh et al. 2017). Additional advantages of assessing fish communities using BRUV systems include their non-destructive nature as a sampling technique (essential for MPA monitoring), the ability to collect highly accurate fish measurements for biomass calculations, the creation of a video record that can be re-analyzed to extract novel information or correct for inter-observer biases, the ability to observe species targeted by fishing and non-targeted species, and the opportunity to collect visual data on specieshabitat associations (Cappo et al. 2007, Langlois et al. 2010, Bennett et al. 2016). BRUV systems have become a widely implemented technique in many parts of the world as a non-destructive, fishery-independent data collection tool for monitoring MPAs (Rees et al. 2004,

Langlois et al. 2006, Kelaher et al. 2014) but are still in the early stages of use in California.

Using BRUV survey data from Carrington Point (Pt.) SMR and Anacapa SMR/SMCA and nearby reference areas, we measured the importance of habitat type, depth, and protection status on the abundance and biomass of targeted fish species. These 2 focal MPAs are nested in larger networks; first, a network within the Channel Islands (Fig. 1B), which itself is nested in a large California statewide network of protected areas. Previous studies have shown individual variation in MPA performance across the network for shallow reefs and kelp forests (Caselle et al. 2015, Ziegler et al. 2023). Here we used 2 MPAs to demonstrate the potential for individual MPA variation in performance in presumably more stable, deeper-water habitats.

2. MATERIALS AND METHODS

2.1. Sampling design

Our BRUV design was modified from existing designs used extensively to monitor a variety of marine habitats in Australia (Willis & Babcock 2000, Goetze et al. 2021, Harvey et al. 2021). These BRUV stereo camera systems consist of a trapezoidal steel frame with 2 GoPro cameras (medium FOV, 30 fps, 1080 p) mounted in a calibrated stereo configuration. Each BRUV unit is equipped with 2 underwater dive lights to augment typically low ambient light conditions and a perforated PVC bait canister located in front of the camera array. To ensure accurate length measurements, we calibrated stereo camera systems before and after each sampling season (Harvey & Shortis 1998).

To standardize sampling location selection, we created a fishnet grid of 100 m × 100 m cells (ArcGIS Pro2.1) applied across each MPA and its respective reference area (the nearest qualifying habitat adjacent to each MP. Grid cells were allocated into 3 depth bins (30-50, 50-70, and 70-100 m), and habitat maps from the California Seafloor Mapping Program (https://www. usqs.gov/centers/pcmsc/science/california-seafloormapping-program) were used to calculate the amount of hard bottom within each grid cell. Grid cells that contained >15% hard bottom for the 2 shallowest depth bins and >5% hard bottom for the deepest depth bin were selected as eligible sampling locations. Deeper grid cells (70–100 m) contained considerably less hard bottom and required the lower criteria to ensure an even number of potential sampling grid cells across the depth bins. Because the Carrington Pt. SMR has very little habitat > 50 m deep, survey depths at this site were generally limited to this maximum depth, while surveys at Anacapa extended to 100 m depth. A map showing each MPA/reference zone polygon can be found in Fig. S1 in the Supplement at www.int-res.com/articles/ suppl/m751p115_supp.pdf.

We selected daily BRUV sampling locations haphazardly from the pool of qualifying grids cells, making sure that surveys were set at a minimum distance of 250 m between units to minimize the effects of bait plume and reduce the likelihood of fish being resampled. We baited BRUVs with whole, moderately scored chub mackerel *Scomber japonicus*, which was replenished before each survey (Dorman et al. 2012, Jones et al. 2020), and deployed each BRUV for a minimum of 30 min (Harasti et al. 2015). Sampling effort included both MPA and reference sites (where fishing is allowed) on any single day to control for environmental variability between sampling days.

Sampling took place from August to October for 4 consecutive years (2019–2022). In 2020, we also sampled in June at Anacapa Island as a response to the shutdown of commercial passenger fishing vessels at that location due to COVID-19 restrictions. We tested for seasonal or COVID-related effects, but because we found none, these data were included in all analyses unless otherwise noted. A map of all sampling locations by year is available in Fig. S2.

2.2. Video processing

We analyzed video files using observation logging and 3D measurement software (SeaGIS Event-Measure; www.seagis.com.au). We identified all observed fishes to the lowest taxonomic group possible. In order to quantify the relative abundance of fishes, we recorded MaxN, the maximum number of individuals of a species present in a single video frame (Willis & Babcock 2000), using the first 30 min of each BRUV survey (Harasti et al. 2015), for every species observed. Without the ability to discern individuals or distinguish multiple sightings of the same fish, MaxN is a conservative estimate of relative abundance that has become the standard metric for BRUV surveys (Langlois et al. 2020).

We also measured the total length for every fish observed in the MaxN video frame. In some instances, we were not able to accurately measure all individuals in a particular MaxN frame due to the camera angle or obstructions blocking one of the cameras. In this case, we measured as many individuals as possible from the MaxN frame. We estimated the biomass of individual fishes using an allometric length-weight conversion: W = aTL, where parameters a and b are species-specific constants, TL is total length in cm, and W is weight in grams. We obtained length-weight fitting parameters from the literature and FishBase (Froese & Pauly 2000, Love 2011). Because not all fish in the MaxN frame can be measured (some individuals are not within view of both cameras) we used the mean of all individual weights in the MaxN frame, multiplied by the MaxN for each individual species to estimate biomass. We classified species as targeted by fishing or nontargeted by fishing using information from the California Department of Fish and Wildlife as well as local knowledge of the authors.

In order to understand the influence of habitat on MPA effectiveness, and because we cannot control the precise view of the forward-facing BRUV cameras, we classified habitat based on substrate characteristics within the field of view of the cameras. The 4 habitat classifications were 'hard', 'mixed hard', 'mixed soft', and 'soft' (categories are defined in Table S1).

To standardize the quality of videos used in analysis, we quantified the 'usability' of each video. All videos were assigned a usability score that considered the amount of visible substrate and video length (Table S2). Any videos in which the BRUV unit did not remain upright for the entire 30 min (i.e. scores of 3 or 4) were removed from analyses.

2.3. Focal fish species

In addition to groupings of all targeted species and all non-targeted species, we also analyzed patterns of biomass and MaxN for specific focal species. Ocean whitefish, the most abundant species observed in our study, are medium-sized (39–56 cm TL at sexual maturity) tilefish (Malacanthidae) common in southern California and Baja, Mexico. Typically residing at the periphery of rocky reefs, ocean whitefish primarily feed on small invertebrates often in nearby sandy flats (Love 2011). This species is a staple for the recreational fishery in the heart of its geographic range and is likely to benefit from fishing protection.

California sheephead, which appeared on nearly half of our surveys, are the largest and only targeted wrasse (Labridae) species inhabiting the study area. California sheephead are not only highly targeted by recreational anglers, but also by a smaller live-fish commercial fishery (Love 2011). This species is a protogynous hermaphrodite (female at birth with the potential to transition to male later in life), and thus fishing pressure has a more complex effect on the sex/size structure and the reproductive potential of local populations (Hamilton et al. 2007).

With 21 unique species of rockfish observed in this study, we more closely examined a sub-group of rockfish that we considered 'targeted' and are more likely to respond to protection from fishing. This grouping excludes small-bodied rockfish that are infrequently caught and almost never retained by anglers, such as halfbanded rockfish Sebastes semicinctus and calico rockfish S. dallii. Three targeted species, namely vermilion rockfish S. miniatus, copper rockfish S. caurinus, and blue rockfish S. mystinus, made up over 70% of the targeted rockfish observed on our surveys. Vermilion and copper rockfish are long-lived (50-60 yr), high-value species that have a significant recreational and commercial fishery, increasing their potential to benefit from protection. Blue rockfish are a schooling, largely planktivorous species that is of moderate value to recreational and commercial fishing (Love 2011).

2.4. Abundance and biomass modeling

Generalized linear models were used to understand how differences in habitat type, depth, and MPA status influenced fish abundance and biomass. To account for species-specific habitat preference and other potential traits (e.g. movement patterns), we ran models on targeted rockfish, ocean whitefish, and California sheephead separately. We also ran separate models for each island because the depth ranges differed (Carrington Pt. SMR: 30–60 m; Anacapa SMCA/SMR: 30–90 m). Predictor variables were MPA status, depth, and habitat type, as well as the interaction between MPA status and habitat type to test if the effect of MPA status is similar across different habitat types:

- Biomass (kq) or Abundance (count) = MPA Status
 - + Depth + Habitat + (MPA Status \times Habitat) (1)

All statistical models and model diagnostics were run using R version 4.3.0 (R Core Team 2023). Data from each species and metric (biomass and MaxN) combination was zero inflated, and the 'fitdistrplus' package (Delignette-Muller & Dutang 2015) and 'descdist()' function were used to determine the best fit distribution. Each data combination was fit to a negative binominal distribution, and the models were run using the 'glm.nb' function in the 'MASS' package (Venables & Ripley 2002). Randomized quantile residuals were calculated and tested for overdispersion using the 'DHARMa' package (Hartig 2022), and all dispersion statistics were <1. The 'visreg' package (Breheny & Burchett 2017) was used to visualize how each model parameter influenced a particular metric and represents each model estimate when all other predictors are held constant. We standardized the visualizations by setting MPA status to 'MPA', habitat type to 'Hard', and depth as the median depth from a particular island. Each regression plot shows a model prediction, 95% confidence interval, and the partial residuals of the model. We calculated the percent deviance explained for each model to understand how well our predictors explained the variation in the data.

2.5. Population size structure

To understand how protection from fishing alters the demographics of targeted fish populations, we created density plots using all measured total lengths of the 3 most common targeted rockfishes (copper, vermillion, and blue rockfish), ocean whitefish, and California sheephead. These curves represent the relative probability of observing an individual fish at a given total length (mm) for each species where the area under each curve is equal to 1.

2.6. Habitat

To compare the overall amount of hard bottom in our depth ranges between the MPA and reference areas, we calculated the number of eligible grid cells within each particular depth bin (30-50, 50-70, and70-100 m). We also report the total study area of each MPA and reference area from which all grid cells, hard and soft, were analyzed. MPA area was calculated from defined MPA boundaries and, for the inshore boundary, the 30 m isobath. Calculating the area of the reference zone was somewhat subjective because the eastern and western boundaries are not designated as they are for the MPA. At Anacapa, we used the 30 and 100 m isobaths for the inshore and offshore boundary, respectively, and for eastern and western boundaries, we estimated the area along the reef contours. At Carrington Pt., we followed a similar protocol, although the northern/offshore boundary was set by the 50 m isobath.

3. RESULTS

We completed 280 and 341 BRUV surveys at Carrington Pt. and Anacapa Island, respectively, for a total of 621 surveys. Of those 621 surveys, 493 (79%) were classified as usable and were included in analyses (Table S3). In general, 2 conditions resulted in unusable video: very strong currents and/or highly rugose habitat causing the BRUV system to not land upright, or to tip over at some point during the video survey.

3.1. Habitat description

We found differences in amount and type of rock at the 2 islands. When rock was present at Anacapa, the habitat was characterized by a patchy reef system, with most videos classified as mixed soft or mixed hard (Fig. 2; Table S4). In addition, the Anacapa MPA contained more soft habitat classifications than the other island/MPA groupings. Carrington Pt. was characterized by more consistent hard bottom, with most videos classified as hard or mixed hard (Fig. 2; Table S4).

Similar to the video observations, benthic habitat maps showed a disparity between the amount of rocky substrate present in the Anacapa MPA and reference area, with 3-5 times more eligible grid cells in the reference area. Although the reference area was twice as large as the MPA, there still appears to be proportionally more abundant hardbottom habitat in the reference area. This is particularly true for the deepest zone (70-100 m), which had 5 times as many eligible grid cells (MPA = 33 vs. reference = 167; Table S5) in the reference area. Carrington Pt. had a relatively similar number of eligible grid cells within the 30-50 m depth bin, with the reference area only being slightly larger (Table S5). Overall, the benthic habitat maps used to identify eligible grid cells generally agreed with the observed habitat data from BRUV surveys, with Carrington Pt. containing more consistent hard bottom and Anacapa containing more sand-rock ecotone habitat.



Fig. 2. Proportion of observed habitat by depth bin from Anacapa and Carrington Point. Habitat was classified from BRUV surveys and was categorized based on visible habitat in which a survey was conducted

3.2. Species observed

We observed a total of 65 fish species on the BRUV surveys across 4 yr and both locations. Total summed MaxN and frequency of occurrence for each species are shown in Table S6. A single species, ocean whitefish, accounted for 32% of the summed MaxN values. These fish had the highest abundance across our study and were present on 84% of all surveys. Two species, copper rockfish and California sheephead, had both high MaxN values and high frequency of occurrence at both islands. Copper rockfish occurred on 48% of surveys while California sheephead occurred on 49% of surveys (Table S6). Four schooling species, namely jack mackerel Trachurus symmetricus, blacksmith Chromis punctipinnis, halfbanded rockfish, and blue rockfish, were abundant when observed but occurred less frequently (4-27% of all surveys).

The BRUVs were able to observe species rarely seen in other surveys in California (e.g. SCUBA, remotely operated vehicle [ROV], submersible). These included species of concern including giant sea bass *Stereolepis gigas* (50 individual observations), bocaccio *Sebastes paucispinis* (79 individuals) and bat rays *Myliobatis californica* (11 individuals) as well as various species of the more cryptic flatfishes (including California halibut *Paralichthys californicus* and Pacific sanddab *Citharichthys sordidus*). Although many species were documented by only a single individual, this type of presence-only data can be useful for documenting climate-induced range shifts, invasions, and habitat associations.

3.3. Abundance and biomass

3.3.1. Targeted rockfish

At both islands, targeted rockfishes showed similar patterns, with habitat type and depth being significant predictors of both biomass and abundance. Generally, both biomass and abundance increased as the availability of hard substrate increased, with hard vs. mixed soft and soft being highly significant (all 8 comparisons across island, metric, hard vs. mixed soft, and hard vs. soft had p-values <0.001; Fig. 3; Table S7). At Anacapa, depth was a significant predictor for targeted rockfish abundance and biomass, with both metrics increasing with depth (MaxN p < 0.001; biomass p < 0.001; Fig. 3; Table S7). The patterns at Carrington Pt. were less consistent, with depth only being a significant predictor of biomass (p = 0.01; Fig. 3; Table S7), most likely related

to the limited depth range at this location compared to Anacapa. Interestingly, the interaction between soft habitat and MPA status was almost significant for targeted rockfish biomass at Anacapa. Here, soft habitat was the only habitat type predicted to have greater targeted rockfish biomass inside the MPA compared to the reference area (p = 0.06, estimate = -1.43; Table S7, Fig. S3A). This interaction term indicates that the model estimate for soft-bottom habitats in the MPA was greater compared to reference areas, while in other habitats (e.g. hard, mixed hard, and mixed soft), the reference area estimate was larger. The only significant MPA effect, beyond the influence of habitat and depth, was for targeted rockfish biomass inside the Carrington Pt. MPA (p = 0.04; Fig. 3; Table S7).

3.3.2. Ocean whitefish

Depth was a strong predictor of ocean whitefish biomass and abundance, with higher abundance and biomass found in shallower waters (p < 0.001 for all comparisons of MaxN and biomass at both islands; Fig. 4; Table S8). The only significant habitat predictor was for soft habitats at Carrington Pt., with both abundance and biomass significantly lower on sandy, softbottom habitats compared to hard substrate (p = 0.05; Fig. 4; Table S8). We did not find a significant interaction term between MPA status and habitat type for either ocean whitefish metric (Fig. S4, Table S8). Anacapa showed a significant MPA effect, with the abundance and biomass of ocean whitefish greater inside the MPA (MaxN p = 0.02; biomass p = 0.01; Fig. 4; Table S8).

3.3.3. California sheephead

Similar to ocean whitefish, depth was a strong predictor of California sheephead biomass and abundance, with higher levels found in the shallower waters (Carrington Pt. and Anacapa MaxN p < 0.001; Carrington Pt. biomass p < 0.01; Anacapa biomass p < 0.001; Fig. 5; Table S9). California sheephead were significantly less abundant on 2 habitat types at Anacapa, mixed soft and soft, compared to hard substrate (mixed soft p = 0.03; soft p < 0.01; Fig. 5; Table S9), and had significantly lower biomass on soft vs. hard habitat at Anacapa (soft p = 0.05; Fig. 5; Table S9). At Carrington Pt., habitat alone was not a significant predictor of sheephead abundance or biomass. We did find a significant interaction term for sheephead bio-



Fig. 3. Partial model predictions from a negative binomial generalized linear model used to test the effect of marine protected area (MPA) status, depth, and habitat on the maximum number of individuals of a species present in a single video frame (MaxN) and biomass of targeted rockfishes for (A) Anacapa and (B) Carrington Point. The blue line and shaded region represent the model predictions and 95% CI, respectively, for a particular predictor variable when all other variables are held constant. The points represent the partial residuals of the model. M H: mixed hard habitats; M S: mixed soft habitats



Fig. 4. As in Fig. 3, but for ocean whitefish



Fig. 5. As in Fig. 3, but for California sheephead

mass and abundance between soft habitat and MPA status at Carrington Pt., but these results had very few observations of sheephead on soft habitat across protection zones, limiting inference from this result (Fig. S5). At Anacapa, we found a significant negative MPA effect for the abundance and biomass of sheephead, with greater values in the reference area (MaxN p = 0.01, estimate = 1.01; biomass p = 0.03, estimate = 1.09; Fig. 5; Table S9).

3.4. Deviance explained

Across all species and both metrics, the model framework performed slightly better at Anacapa compared to Carrington Pt., with the deviance explained ranging from 0.46 to 0.22 at Anacapa and from 0.22 to 0.17 at Carrington Pt. (Tables S7–S9). This is likely related to the larger available depth range at Anacapa and that depth was a significant predictor of abundance and biomass for all 3 species in this study. The highest deviance explained (DE) was for California sheephead at Anacapa (DE = 0.46 for MaxN; DE = 0.45 for Biomass; Table S9) and is most likely related to the smaller range in MaxN and biomass values associated with the California sheephead data compared to the larger range associated with targeted rockfish and ocean whitefish.

3.5. Focal species size structure

We plotted the density of individual fish total lengths in MPAs and reference areas for the 5 most common targeted species in our study; copper rockfish (n = 506), vermilion rockfish (n = 459), blue rockfish (n = 1760), ocean whitefish (n = 5767), and California sheephead (n = 603) (Fig. 6). Copper rockfish were generally larger inside the MPAs at both Anacapa and Carrington Pt., with the MPA containing more individuals >400 mm TL (Fig. 6A). Vermilion rockfish had a bimodal size structure,



Fig. 6. Species-specific densities of the 5 targeted fish species: (A) copper rockfish, (B) vermillion rockfish, (C) blue rockfish, (D) ocean whitefish, and (E) California sheephead from Anacapa and Carrington Point

with multiple size classes present at Carrington Pt. and Anacapa (Fig. 6B). Inside the Anacapa MPA, the population size structure was dominated by smaller individuals (<20 cm), with proportionally fewer adult fishes, while the population size structure patterns for vermillion rockfish at Carrington Pt. appeared to be more related to protection from fishing, with proportionally larger individuals inside the MPA. The smallerbodied schooling blue rockfish had similar population size structure across protection zones at Carrington Pt. and Anacapa (Fig. 6C). Ocean whitefish and California sheephead showed similar patterns at Anacapa and Carrington, with a greater proportion of larger individuals inside the MPA at each location (Fig. 6D,E).

4. DISCUSSION

In this study, we show that environmental drivers, such as depth and habitat availability, need to be considered when assessing the ability of MPAs to enhance the biomass and abundance of targeted species. MPA effectiveness, in both tropical and temperate marine ecosystems, has been linked to benthic features such as the amount of consolidated hard bottom, reef complexity, and habitat continuity (Dames et al. 2020). Individual species have been shown to respond differently to such features, and hence species-habitat associations must be considered during MPA implementation, when defining MPA goals, and when assessing performance. A lack of preferred habitat inside the protection zone can limit the ability of MPAs to enhance species and functional diversity (Navarro-Martínez et al. 2023). MPAs are designed as ecosystem management tools, with broad goals of protecting biodiversity, yet individual species can have widely varying habitat requirements. Many MPA design processes, especially for MPA networks, try to encompass habitat replication and representation (Roberts et al. 2003, Halpern et al. 2010, Saarman et al. 2013), but in practice, it is impossible to design optimally for every species. Thus, accounting for habitat in post implementation monitoring and choice of reference areas can help to set realistic expectations for MPA performance.

The Carrington Pt. MPA and adjacent reference area had similar amounts of continuous hard substrate, which indicated that the greater biomass of targeted rockfishes in the MPA was likely related to protection more than habitat. Rockfishes (genus *Sebastes*) are generally characterized as demersal, rock-associated, and heavily fished, and it appears that the Carrington Pt. MPA is well designed for their protection. Inside the Anacapa MPA, there was a notable lack of continuous hard substrate, particularly at depth, which appeared to limit the performance of the MPA at that location for this rock-associated fish group and makes comparisons with the paired reference area challenging. However, the Anacapa MPA is likely more effective at enhancing the ocean whitefish population, a more mobile species that is associated with the sand-rock ecotone (Bellquist et al. 2008). Habitat maps and in situ observations from the Anacapa MPA and reference area showed that these locations were dominated by a mix of soft, sandy habitat and isolated patch reefs. Ocean whitefish abundance and biomass was also strongly associated with depth, but unlike targeted rockfish, ocean whitefish are more abundant in shallower areas, with the majority of observations occurring shallower than 60 m.

The examples above, comparing very different fish species groups, demonstrate how scientific guidelines and a network approach to MPA design can help meet ecosystem goals and protect populations of different species across a variety of habitats. The creation of networks, with numerous reserves spread across various habitats, has become the standard by which to structure marine no-take zones, both at the local and global scale. The California Marine Life Protection Act (MLPA) established the network of MPAs throughout California with the goals of protecting ecosystem biodiversity and protecting and rebuilding populations of key species. This process relied on MPA design guidelines from a scientific advisory committee, and chief among those guidelines was 'habitat representation' throughout the network (Saarman et al. 2013). At the network level, this was achieved by including a variety of major habitat types, such as rocky intertidal zones, estuaries, and shallow and deep subtidal habitat. Here we show that a more nuanced treatment of habitat within an MPA can provide critical information on individual MPA performance for key species (Young & Carr 2015).

By incorporating habitat availability in and out of the MPAs and studying deeper zones than previously explored, we found several results that differed from prior studies and/or our expectations. Contrary to our predictions and prior studies of shallower reefs in the area (Caselle et al. 2015, Honeyman et al. 2023), California sheephead abundance and biomass at these deeper depths was significantly greater in the Anacapa reference area, which is open to fishing. Active tracking has shown that California sheephead use a variety of habitats, but tend to favor rocky reefs, spending 50–70% of their time on hard, consolidated substrate (Topping et al. 2005). The deepwater habitat within the Anacapa MPA lacks this hard substrate, which is more abundant in the shallow kelp forests of the Anacapa MPA. This illustrates the importance of surveying the full range of depths for species of interest when evaluating MPA performance or advising fisheries management. A more comprehensive approach to MPA design and management can be achieved by filling gaps in our knowledge of how MPAs may differentially enhance fish communities at all depths and habitat types.

This study, much like other MPA evaluations, does have known limitations. To measure the ecological effects of an MPA, especially without data for that site prior to MPA implementation, researchers often use a nearby fished area (reference site) as a control by which hypotheses about the cessation of fishing can be tested. Selection of these 'control' sites must weigh numerous factors, such as habitat type, depth range, species composition, and seasonal oceanographic characteristics, when deciding on location and scale. Often the easiest way to broadly control for various environmental factors is by placing the reference area in close proximity to the MPA. However, unlike strictly controlled lab experiments, MPAs are expected to affect control areas by design. Adjacent placement of reference areas, while often meeting goals of controlling for environmental conditions and perhaps even habitat, creates a scientific design challenge, with the likelihood that reference sites will be affected by the presence of a nearby MPA through the redistribution of fishing effort, and potential 'spillover' of fish coming from the MPA (Caselle et al. 2015, Di Lorenzo et al. 2016).

Anacapa Island presents an interesting case study in reference site selection. Previous studies of very deep water habitats using ROVs excluded the Anacapa MPA from analyses, citing a lack of an appropriate reference area (Karpov et al. 2012). This exclusion comes despite containing one of the oldest MPAs in the Northern Channel Islands network, with the Anacapa eastern SMR established in 1978. Here we chose the reference location as immediately offshore of, and deeper than, long-standing reference sites used for shallower rocky reef and kelp forest studies (Caselle et al. 2015, White et al. 2021). While shown to be appropriate for shallow rock reef, we found that in deeper waters, the reference area contained more deep rock than in the MPA, potentially limiting our ability to detect an MPA effect on rock-associated species (e.g. targeted rockfish, California sheephead). However, both reference area and MPA contained similar amounts of deep sand-rock ecotone habitat favored by ocean whitefish, the most numerous species of fish in our study. Hence tradeoffs exist where reference site selection must incorporate species habitat associations, assess MPA goals, work to inform management decisions, and be beholden to the realities of field studies.

A consistent MPA effect was observed in the size structure of 3 targeted rockfish species, ocean whitefish, and California sheephead, with the greater proportion of larger individuals inside the protection zone. This is not surprising as one of the earliest effects of a cessation of fishing is to allow larger individuals to grow and persist (Taylor & McIlwain 2010, Bejarano et al. 2019, Mumby et al. 2021). The notable exception was for vermillion rockfish inside the Anacapa MPA, which contained fewer large individuals than the reference area. Interestingly, the size structure for vermilion rockfish at both study sites revealed distinct cohorts at 2 size classes. This likely indicates periodic strong recruitment/survivorship for this species and provides an opportunity to track these cohorts over time. Repeated annual sampling such as done here allows the tracking of these types of population demographics through time to better understand year to year variability in recruitment dynamics for valuable fishery species. The ability to collect size structure data, with tools such as stereo BRUVs, can provide important additional insight into the effects of protection on these populations and is particularly important for the management of long-lived species like rockfishes. Although we did not directly analyze annual changes in size class and recruitment here, stereo-video surveys are well suited for such analyses and are a potential avenue for future work.

The ability of MPAs to conserve and rebuild fish communities has been shown to improve with the protection of habitat across a wide range of depths (Curley et al. 2002, Goetze et al. 2021). Many species of fish utilize different depth zones and habitats across life stages, often with juvenile fish recruiting to relatively protected shallow nursery habitats and moving to deeper offshore habitats as they mature (Gibson et al. 2002, Aburto-Oropeza et al. 2009, Li et al. 2022). This ontogenetic movement is common with a number of the rockfish species observed in this study (Love et al. 1991) and should be considered in the context of MPA performance. The availability of suitable habitat along the ontogenetic depth profile likely influences the distribution of rockfish at varying life stages. Additionally, seasonal spawning migrations across depth gradients are observed globally and emphasize the significance of providing protection across this gradient (Lombardo et al. 2020, Luo et al. 2020, Thorburn et al. 2021). We observed a distinct break in suitable

rockfish habitat within the Anacapa MPA complex, with very little deep habitat (>70 m) present. Not only does this reduce the overall available habitat for adult rockfish, but it likely also reduces the potential for this MPA to provide a valuable ontogenetic bridge for species with high habitat affinity. An MPA that contains contiguous habitats within the full depth range of a species is better equipped to protect that species at all stages of life, enhancing the benefits experienced by protection from fishing.

The protection of connected depth gradients is also likely to provide resilience for fish populations faced with environmental stressors including seasonal changes in water temperature, short-term temperature anomalies (marine heat waves), and long-term changes resulting from global climate change. In an ever-warming global ocean, it has become necessary for fish communities to adapt to rising sea temperatures, often through range shifts in latitude or depth (Perry et al. 2005, Chaikin et al. 2022, Dahms & Killen 2023). Access to deep refuge habitat can improve the resilience of some fish communities and allow them to survive shallow water stressors (MacDonald et al. 2016, Pereira et al. 2018). Inversely, global climate change has been shown to reduce deep-water dissolved oxygen, creating upward pressure on fish communities escaping oxygen minimum zones (OMZs) (Ross et al. 2020). This has been observed around Southern California's Channel Islands, with some rockfish communities shifting to shallower depths as a response to OMZ shoaling (Meyer-Gutbrod et al. 2021). Spatial protections such as MPAs can provide corridors of protected habitat across depth zones to allow for this type of adaptive migratory process. The combined effects of large-scale environmental stressors and species-specific life histories on habitat use across depth gradients have the potential to alter the effectiveness of an MPA in the near and long term.

We used the same model framework across 2 species and 1 species group to understand the importance of MPA status, habitat type, and depth in determining patterns of abundance and biomass. Across all models, the model performance indicated that these factors were important in explaining the observed patterns; however, other aspects of the system could be incorporated to more accurately evaluate conservation efforts. Future studies will work to integrate factors such as distance to reserve edge, presence of other ephemeral habitats (e.g. ophiuroid or urchin barrens), and oceanographic parameters. BRUVs are particularly well suited to adaptive monitoring programs, in that they are deployable in multiple habitats, are relatively inexpensive to operate, and provide a video record that can be reassessed over time (Langlois et al. 2010, Whitmarsh et al. 2017).

The California MPA network will continue to work towards protecting marine biodiversity and populations of key fished species across the state. Our results demonstrate the realized benefits of a well designed and well enforced protection network, with a nuanced look at how 2 geographically distinct MPAs might effectively protect different fisheries targets. We also highlight the importance of acknowledging and describing the limitations of particular MPAs when assessing an MPA network to align expectations with realistic outcomes. Continued monitoring efforts will provide valuable ecological knowledge as MPAs age and management efforts adapt to preserve fisheries in the face of climate uncertainty.

Acknowledgements. This work would not be possible without funding support from the California Ocean Protection Council and California Seagrant (Grant Numbers R/MPA-43 and R/MPA-48). Special thanks to C. Honeyman, A. Parsons-Field, J. Kolda, B. Brock, B. Behar, P. Campbell, W. Horstmeyer, N. Castaneda, S. Jaeger, K. McCaffrey, S. Urgoiti Crespo, G. Hansen, J. Harding, and M. O'Connell for assistance with fieldwork. Thank you to Y. Chen, Y. Sha, T. Hozdic, G. Simmons, J. Zounes, and J. Eisaguirre for video analysis; S. Gaines, D. McCauley, R. Starr, J. Lindholm, and J. Todd for advice and insightful discussions; and A. Giraldo Ospina and C. Free for help with data analysis.

LITERATURE CITED

- Aburto-Oropeza O, Dominguez-Guerrero I, Cota-Nieto J, Plomozo-Lugo T (2009) Recruitment and ontogenetic habitat shifts of the yellow snapper (*Lutjanus argentiventris*) in the Gulf of California. Mar Biol 156:2461–2472
- Andradi-Brown DA, Macaya-Solis C, Exton DA, Gress E, Wright G, Rogers AD (2016) Assessing Caribbean shallow and mesophotic reef fish communities using baitedremote underwater video (BRUV) and diver-operated video (DOV) survey techniques. PLOS ONE 11:e0168235
- Baillon S, Hamel JF, Wareham VE, Mercier A (2012) Deep cold-water corals as nurseries for fish larvae. Front Ecol Environ 10:351–356
- Bejarano S, Pardede S, Campbell SJ, Hoey AS, Ferse SCA (2019) Herbivorous fish rise as a destructive fishing practice falls in an Indonesian marine national park. Ecol Appl 29:e01981
- Bell JJ, Micaroni V, Harris B, Strano F, Broadribb M, Rogers A (2024) Global status, impacts, and management of rocky temperate mesophotic ecosystems. Conserv Biol 38:e13945
- Bellquist LF, Lowe CG, Caselle JE (2008) Fine-scale movement patterns, site fidelity, and habitat selection of ocean whitefish (*Caulolatilus princeps*). Fish Res 91:325–335
- Bennett K, Wilson SK, Shedrawi G, McLean DL, Langlois TJ (2016) Can diver operated stereo-video surveys for fish be used to collect meaningful data on benthic coral reef communities? Limnol Oceanogr Methods 14:874–885

- Breheny P, Burchett W (2017) Visualization of regression models using visreg. R J 9:56–71
- Brokovich E, Einbinder S, Shashar N, Kiflawi M, Kark S (2008) Descending to the twilight-zone: changes in coral reef fish assemblages along a depth gradient down to 65 m. Mar Ecol Prog Ser 371:253–262
- Brown K, Monk J, Williams J, Carroll A, Harasti D, Barrett N (2022) Depth and benthic habitat influence shallow and mesophotic predatory fishes on a remote, high-latitude coral reef. PLOS ONE 17:e0265067
- California Department of Fish and Game Marine Region (2003) Review of some California fisheries for 2002: market squid, sea urchin, Dungeness crab, prawn, coastal pelagic finfish, albacore, ocean salmon, nearshore livefish, Pacific herring, and recreational. CCOFI Rep 44: 10-27
- Cappo M, Speare P, De'ath G (2004) Comparison of baited remote underwater video stations (BRUVS) and prawn (shrimp) trawls for assessments of fish biodiversity in inter-reefal areas of the Great Barrier Reef Marine Park. J Exp Mar Biol Ecol 302:123–152
- Cappo M, Harvey E, Shortis M (2007) Counting and measuring fish with baited video techniques — an overview. In: Lyle JM, Furlani DM, Buxton CD (eds) Cutting-edge technologies in fish and fisheries science. Australian Society for Fish Biology Workshop Proceedings, Hobart, 28–29 August 2006. Australian Society for Fish Biology, Hobart, p 101–114
- Caselle JE, Rassweiler A, Hamilton SL, Warner RR (2015) Recovery trajectories of kelp forest animals are rapid yet spatially variable across a network of temperate marine protected areas. Sci Rep 5:14102
- Cerrano C, Bastari A, Calcinai B, Di Camillo C and others (2019) Temperate mesophotic ecosystems: gaps and perspectives of an emerging conservation challenge for the Mediterranean Sea. Eur Zool J 86:370–388
- Chaikin S, Dubiner S, Belmaker J (2022) Cold-water species deepen to escape warm water temperatures. Glob Ecol Biogeogr 31:75–88
- Curley BG, Kingsford MJ, Gillanders BM (2002) Spatial and habitat-related patterns of temperate reef fish assemblages: implications for the design of Marine Protected Areas. Mar Freshw Res 53:1197–1210
- Dahms C, Killen SS (2023) Temperature change effects on marine fish range shifts: a meta-analysis of ecological and methodological predictors. Glob Change Biol 29: 4459–4479
- Dames V, Bernard A, Floros C, Mann B and others (2020) Zonation and reef size significantly influence fish population structure in an established marine protected area, iSimangaliso Wetland Park, South Africa. Ocean Coast Manage 185:105040
- Delignette-Muller ML, Dutang C (2015) fitdistrplus: an R package for fitting distributions. J Stat Softw 64(4):1–34
- Dennis D, Plagányi É, Van Putten I, Hutton T, Pascoe S (2015) Cost benefit of fishery-independent surveys: Are they worth the money? Mar Policy 58:108–115
- Di Lorenzo M, Claudet J, Guidetti P (2016) Spillover from marine protected areas to adjacent fisheries has an ecological and a fishery component. J Nat Conserv 32:62–66
- Dorman SR, Harvey ES, Newman SJ (2012) Bait effects in sampling coral reef fish assemblages with stereo-BRUVs. PLOS ONE 7:e41538
- Field SA, O'Connor PJ, Tyre AJ, Possingham HP (2007) Making monitoring meaningful. Austral Ecol 32:485–491

- Froese R, Pauly D (eds) (2000) FishBase 2000: concepts, designs and data sources. Internation Center for Living Aquatic Resources Management (ICLARM), Los Baños
- Gaines SD, White C, Carr MH, Palumbi SR (2010) Designing marine reserve networks for both conservation and fisheries management. Proc Natl Acad Sci USA 107: 18286–18293
- Gibson RN, Robb L, Wennhage H, Burrows MT (2002) Ontogenetic changes in depth distribution of juvenile flatfishes in relation to predation risk and temperature on a shallow-water nursery ground. Mar Ecol Prog Ser 229: 233–244
- Giraldo-Ospina A, Kendrick GA, Hovey RK (2020) Depth moderates loss of marine foundation species after an extreme marine heatwave: Could deep temperate reefs act as a refuge? Proc R Soc B 287:20200709
- Gleason M, Fox E, Ashcraft S, Vasques J and others (2013) Designing a network of marine protected areas in California: achievements, costs, lessons learned, and challenges ahead. Ocean Coast Manag 74:90–101
- Goetze JS, Wilson S, Radford B, Fisher R and others (2021) Increased connectivity and depth improve the effectiveness of marine reserves. Glob Change Biol 27:3432–3447
- Grorud-Colvert K, Claudet J, Tissot BN, Caselle JE and others (2014) Marine protected area networks: assessing whether the whole is greater than the sum of its parts. PLOS ONE 9:e102298
- Grüss A, Robinson J, Heppell SS, Heppell SA, Semmens BX (2014) Conservation and fisheries effects of spawning aggregation marine protected areas: what we know, where we should go, and what we need to get there. ICES J Mar Sci 71:1515–1534
- Halpern BS, Lester SE, McLeod KL (2010) Placing marine protected areas onto the ecosystem-based management seascape. Proc Natl Acad Sci USA 107:18312–18317
- Hamilton SL, Caselle JE, Standish JD, Schroeder DM, Love MS, Rosales-Casian JA, Sosa-Nishizaki O (2007) Sizeselective harvesting alters life histories of a temperate sex-changing fish. Ecol Appl 17:2268–2280
- Hamilton SL, Caselle JE, Malone DP, Carr MH (2010) Incorporating biogeography into evaluations of the Channel Islands marine reserve network. Proc Natl Acad Sci USA 107:18272–18277
- Harasti D, Malcolm H, Gallen C, Coleman MA, Jordan A, Knott NA (2015) Appropriate set times to represent patterns of rocky reef fishes using baited video. J Exp Mar Biol Ecol 463:173–180
- Harms S, Winant CD (1998) Characteristic patterns of the circulation in the Santa Barbara Channel. J Geophys Res Oceans 103:3041–3065
- Hartig F (2022) DHARMa: residual diagnostics for hierarchical (multi-level/mixed) regression models. R package version 0.4.6. https://CRAN.R-project.org/web/pack ages/DHARMa/index.html
 - Harvey ES, Shortis MR (1998) Calibration stability of an underwater stereo-video system: implications for measurement accuracy and precision. Mar Technol Soc J 32: 3–17
- Harvey ES, McLean DL, Goetze JS, Saunders BJ and others (2021) The BRUVs workshop—an Australia-wide synthesis of baited remote underwater video data to answer broad-scale ecological questions about fish, sharks and rays. Mar Policy 127:104430
- Heyns-Veale ER, Bernard ATF, Richoux NB, Parker D, Langlois TJ, Harvey ES, Götz A (2016) Depth and habitat

determine assemblage structure of South Africa's warm-temperate reef fish. Mar Biol 163:158

- Honeyman C, Carlson P, Jainese C, Parsons-Field A and others (2023) Correspondence among multiple methods provides confidence when measuring marine protected area effects for species and assemblages. J Appl Ecol 60: 2699–2712
- Horn MH, Allen LG (1978) A distributional analysis of California coastal marine fishes. J Biogeogr 5:23–42
- Howard RA, Ciannelli L, Wakefield WW, Haltuch MA (2023) Comparing fishery-independent and fishery-dependent data for analysis of the distributions of Oregon shelf groundfishes. Fish Res 258:106553
- Jacquemont J, Loiseau C, Tornabene L, Claudet J (2024) 3D ocean assessments reveal that fisheries reach deep but marine protection remains shallow. Nat Commun 15:4027
- James LC, Marzloff MP, Barrett N, Friedman A, Johnson CR (2017) Changes in deep reef benthic community composition across a latitudinal and environmental gradient in temperate Eastern Australia. Mar Ecol Prog Ser 565:35–52
- Jones RE, Griffin RA, Januchowski-Hartley SR, Unsworth RKF (2020) The influence of bait on remote underwater video observations in shallow-water coastal environments associated with the North-Eastern Atlantic. PeerJ 8:e9744
- Kahng S, Copus JM, Wagner D (2017) Mesophotic coral ecosystems. In: Rossi S, Bramanti L, Gori A, Orejas C (eds) Marine animal forests: the ecology of benthic biodiversity hotspots. Springer, Cham, p 185–206
- Karpov KA, Bergen M, Geibel JJ (2012) Monitoring fish in California Channel Islands marine protected areas with a remotely operated vehicle: the first five years. Mar Ecol Prog Ser 453:159–172
- Kelaher BP, Coleman MA, Broad A, Rees MJ, Jordan A, Davis AR (2014) Changes in fish assemblages following the establishment of a network of no-take marine reserves and partially-protected areas. PLOS ONE 9:e85825
- ^{*}Langlois T, Chabanet P, Pelletier D, Harvey E (2006) Baited underwater video for assessing reef fish populations in marine reserves. SPC Fish Newsl 118:53–57
- Langlois TJ, Harvey ES, Fitzpatrick B, Meeuwig JJ, Shedrawi G, Watson DL (2010) Cost-efficient sampling of fish assemblages: comparison of baited video stations and diver video transects. Aquat Biol 9:155–168
- Langlois T, Goetze J, Bond T, Monk J and others (2020) A field and video annotation guide for baited remote underwater stereo-video surveys of demersal fish assemblages. Methods Ecol Evol 11:1401–1409
- Lenihan HS, Fitzgerald SP, Reed DC, Hofmeister JKK, Stier AC (2022) Increasing spillover enhances southern California spiny lobster catch along marine reserve borders. Ecosphere 13:e4110
- Li L, Hollowed AB, Cokelet ED, McClure MM, Keller AA, Barbeaux SJ, Palsson WA (2022) Three-dimensional ontogenetic shifts of groundfish in the Northeast Pacific. Fish Fish 23:1221–1239
- ^{*} Lombardo SM, Adams AJ, Danylchuk AJ, Luck CA, Ajemian MJ (2020) Novel deep-water spawning patterns of bonefish (*Albula vulpes*), a shallow water fish. Mar Biol 167:187
 - Love MS (2011) Certainly more than you want to know about the fishes of the Pacific Coast: a postmodern experience. Really Big Press, Santa Barbara, CA
 - Love MS, Yoklavich MM (2006) Deep rock habitats. In: Allen LG, Pondella DJ, Horn MH (eds) The ecology of marine fishes: California and adjacent waters. University of California Press, Berkeley, CA, p 253–266

- Love MS, Carr MH, Haldorson LJ (1991) The ecology of substrate-associated juveniles of the genus Sebastes. Environ Biol Fishes 30:225–243
 - Love MS, Yoklavich M, Thorsteinson LK (2002) The rockfishes of the northeast Pacific. University of California Press, Berkeley, CA
- Luo J, Ault JS, Ungar BT, Smith SG and others (2020) Migrations and movements of Atlantic tarpon revealed by two decades of satellite tagging. Fish Fish 21:290–318
- MacDonald C, Bridge TCL, Jones GP (2016) Depth, bay position and habitat structure as determinants of coral reeffish distributions: Are deep reefs a potential refuge? Mar Ecol Prog Ser 561:217–231
- Maestro M, Pérez-Cayeiro ML, Chica-Ruiz JA, Reyes H (2019) Marine protected areas in the 21st century: current situation and trends. Ocean Coast Manage 171:28–36
- Meyer-Gutbrod E, Kui L, Miller R, Nishimoto M, Snook L, Love M (2021) Moving on up: vertical distribution shifts in rocky reef fish species during climate-driven decline in dissolved oxygen from 1995 to 2009. Glob Change Biol 27:6280–6293
- Miller RR, Field JC, Santora JA, Monk MH, Kosaka R, Thomson C (2017) Spatial valuation of California marine fisheries as an ecosystem service. Can J Fish Aquat Sci 74: 1732–1748
- Moffitt EA, White JW, Botsford LW (2011) The utility and limitations of size and spacing guidelines for designing marine protected area (MPA) networks. Biol Conserv 144: 306–318
- Mumby PJ, Steneck RS, Roff G, Paul VJ (2021) Marine reserves, fisheries ban, and 20 years of positive change in a coral reef ecosystem. Conserv Biol 35:1473–1483
- Murray SN, Ambrose RF, Bohnsack JA, Botsford LW and others (1999) No-take reserve networks: sustaining fishery populations and marine ecosystems. Fisheries 24: 11–25
- Navarro-Martínez ZM, de la Guardia E, Szelistowski WA, Angulo-Valdés JA (2023) Habitats diversity and MPA regulations are insufficient in promoting healthy coral reef fish assemblages in Punta Francés National Park (Cuba). Reg Stud Mar Sci 60:102826
- Noble-James T, Bullimore R, McBreen F, O'Connor J and others (2023) Monitoring benthic habitats in English Marine Protected Areas: lessons learned, challenges and future directions. Mar Policy 157:105852
- Olson AM, Hessing-Lewis M, Haggarty D, Juanes F (2019) Nearshore seascape connectivity enhances seagrass meadow nursery function. Ecol Appl 29:e01897
- Pereira PHC, Macedo CH, Nunes JACC, Marangoni LFB, Bianchini A (2018) Effects of depth on reef fish communities: insights of a 'deep refuge hypothesis' from Southwestern Atlantic reefs. PLOS ONE 13:e0203072
- Perry AL, Low PJ, Ellis JR, Reynolds JD (2005) Climate change and distribution shifts in marine fishes. Science 308:1912-1915
- Ponti M, Turicchia E, Ferro F, Cerrano C, Abbiati M (2018) The understorey of gorgonian forests in mesophotic temperate reefs. Aquat Conserv 28:1153–1166
 - R Core Team (2023) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna
- Rees M, Heyward A, Cappo M, Speare P, Smith L (2004) Ningaloo Marine Park — Initial survey of seabed biodiversity in intermediate and deeper waters (March 2004). Australian Institute of Marine Science, Cape Cleveland

- Roberts CM, Andelman S, Branch G, Bustamante RH and others (2003) Ecological criteria for evaluating candidate sites for marine reserves. Ecol Appl 13:199–214
- Ross T, Du Preez C, Ianson D (2020) Rapid deep ocean deoxygenation and acidification threaten life on Northeast Pacific seamounts. Glob Change Biol 26:6424–6444
- Saarman E, Gleason M, Ugoretz J, Airamé S and others (2013) The role of science in supporting marine protected area network planning and design in California. Ocean Coast Manage 74:45–56
- Sala E, Giakoumi S (2018) No-take marine reserves are the most effective protected areas in the ocean. ICES J Mar Sci 75:1166–1168
- Scholz A, Steinback C, Mertens M, Birney K and others (2006) Commercial fishing grounds and their relative importance off the Central Coast of California. Report submitted to the California Marine Life Protection Act Initiative. Ecotrust, Portland, OR
- Sullivan-Stack J, Aburto-Oropeza O, Brooks CM, Cabral RB and others (2022) A scientific synthesis of marine protected areas in the United States: status and recommendations. Front Mar Sci 9:849927
- Swadling DS, Knott NA, Taylor MD, Coleman MA, Davis AR, Rees MJ (2022) Seascape connectivity of temperate fishes between estuarine nursery areas and open coastal reefs. J Appl Ecol 59:1406–1416
- Taylor BM, McIlwain JL (2010) Beyond abundance and biomass: effects of marine protected areas on the demography of a highly exploited reef fish. Mar Ecol Prog Ser 411: 243–258
- Thorburn J, Wright PJ, Lavender E, Dodd J and others

Editorial responsibility: Elliott Hazen, Pacific Grove, California, USA Reviewed by: T. Langlois and 2 anonymous referees (2021) Seasonal and ontogenetic variation in depth use by a critically endangered benthic elasmobranch and its implications for spatial management. Front Mar Sci 8: 656368

- Topping DT, Lowe CG, Caselle JE (2005) Home range and habitat utilization of adult California sheephead, Semicossyphus pulcher (Labridae), in a temperate no-take marine reserve. Mar Biol 147:301–311
- Venables WN, Ripley BD (2002) Modern applied statistics with S, 4th edn. Springer, New York, NY
- White JW, Yamane MT, Nickols KJ, Caselle JE (2021) Analysis of fish population size distributions confirms cessation of fishing in marine protected areas. Conserv Lett 14: e12775
- Whitmarsh SK, Fairweather PG, Huveneers C (2017) What is Big BRUVver up to? Methods and uses of baited underwater video. Rev Fish Biol Fish 27:53–73
- Williams J, Jordan A, Harasti D, Davies P, Ingleton T (2019) Taking a deeper look: quantifying the differences in fish assemblages between shallow and mesophotic temperate rocky reefs. PLOS ONE 14:e0206778
- Willis TJ, Babcock RC (2000) A baited underwater video system for the determination of relative density of carnivorous reef fish. Mar Freshw Res 51:755–763
- Young M, Carr M (2015) Assessment of habitat representation across a network of marine protected areas with implications for the spatial design of monitoring. PLOS ONE 10:e0116200
- ^SZiegler SL, Johnson JM, Brooks RO, Johnston EM and others (2023) Marine protected areas, marine heatwaves, and the resilience of nearshore fish communities. Sci Rep 13:1405

Submitted: July 16, 2024 Accepted: October 30, 2024 Proofs received from author(s): December 14, 2024